



# MODEL GENERATION TECHNICAL REPORT

## Targeted transgenesis of pCAG- Cre-2A-Flpo-2A-eGFP in HPRT

Project code: R5 / IR3160

Report updated: 07/02/2023

1 PROJECT PROCESS &  
QUALITY CONTROL

2 GENETIC STRATEGY

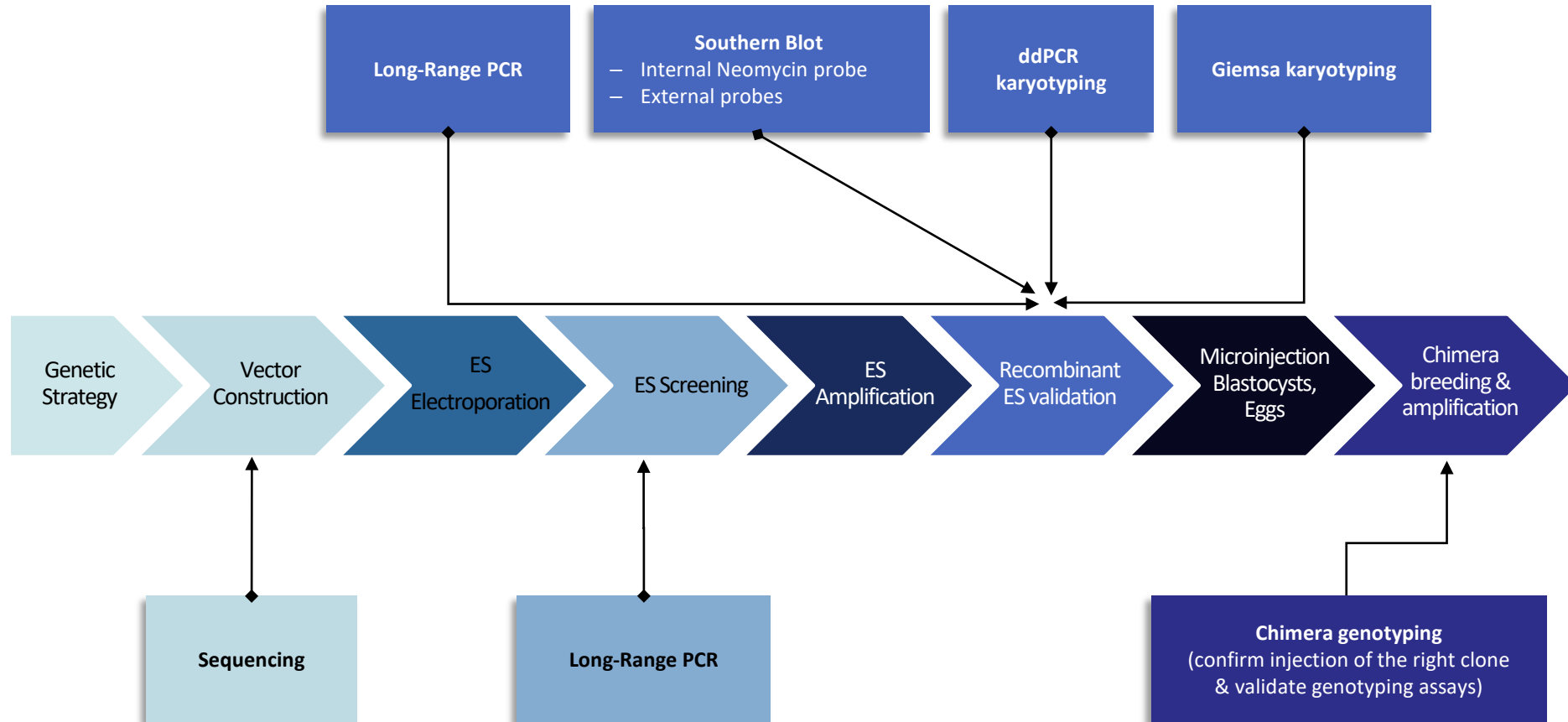
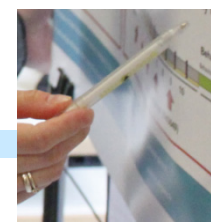
3 HOMOLOGOUS RECOMBINATION  
VECTOR CONSTRUCTION

4 ES ELECTROPORATION & SCREENING OF  
RECOMBINANT CLONES

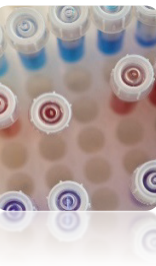
5 MICROINJECTION & BREEDING

6 SEQUENCE OF THE DELIVERED ALLELE

# Project process & quality controls



## 2 GENETIC STRATEGY

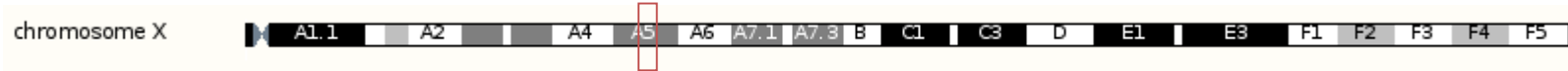


- Target locus structure
- Genetic strategy

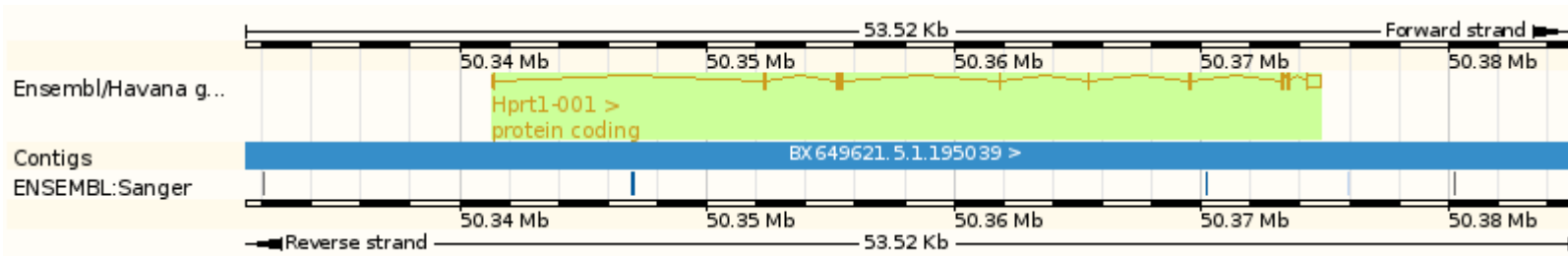
# ■ HPRT mouse genomic locus – structure



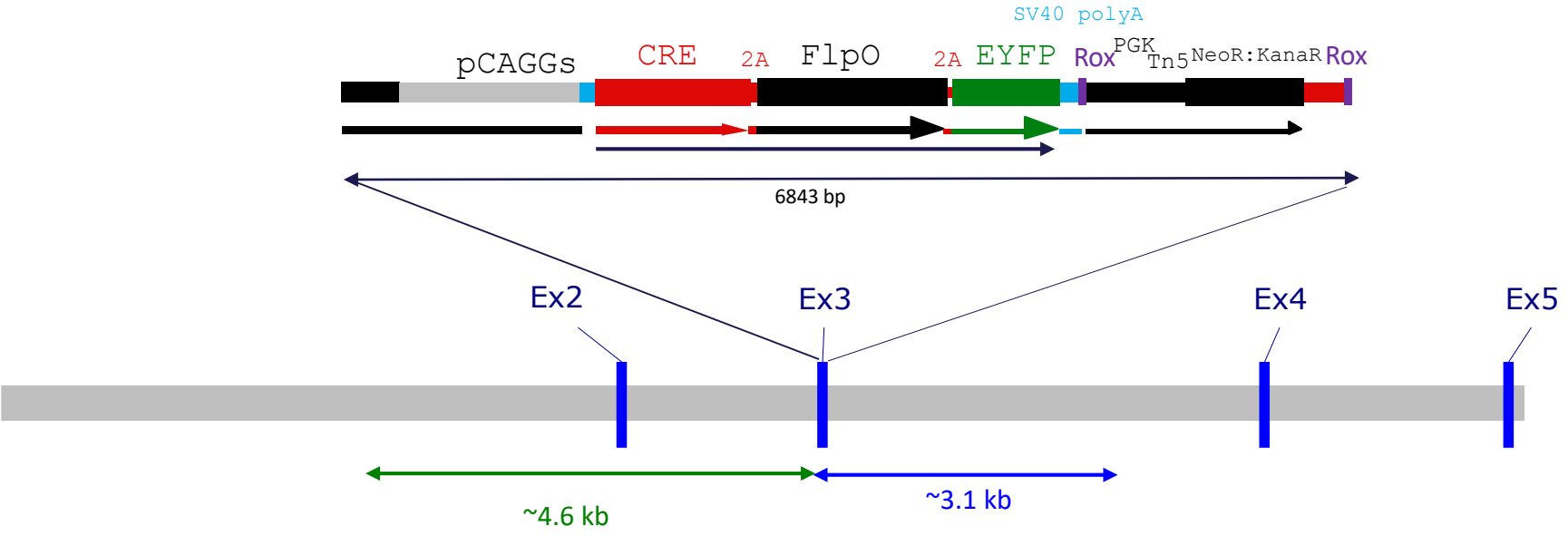
## Location:



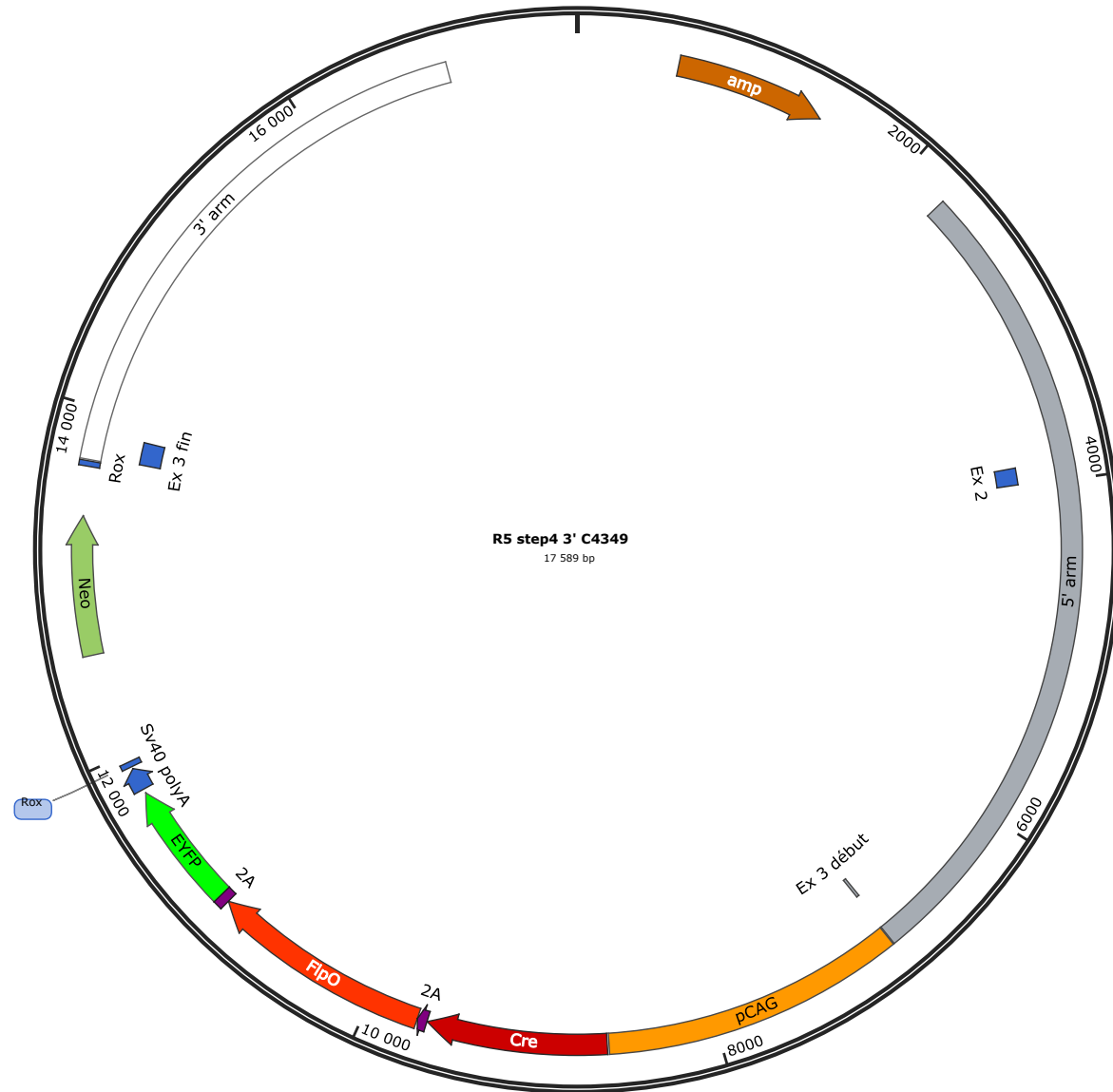
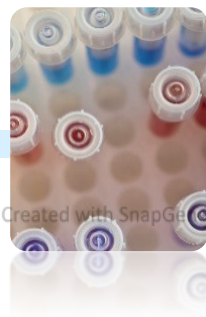
## Ensembl ID: ENSMUSG00000025630



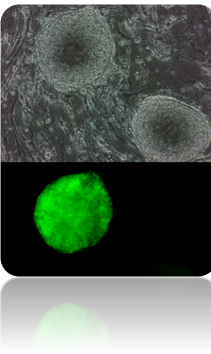
# Strategy: targeted transgenesis of pCAG-CRE-2A-Flpo-2A-eYFP in HPRT



# 3 HOMOLOGOUS RECOMBINATION - VECTOR CONSTRUCTION



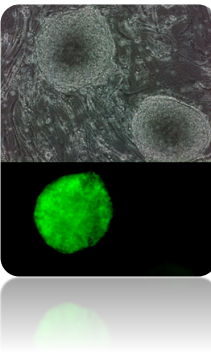
## 4 ES cell electroporation & Screening of recombinant clones



- Electroporation and screening process
- Long range PCR screening – strategy
- Recombinant ES validation by Long Range PCR
- Recombinant ES clones validation by Southern Blot – internal probe
- Recombinant ES clones validation by Southern Blot – External probe
- Aneuploidy screening in ES recombinant clones



## ■ Electroporation and screening process



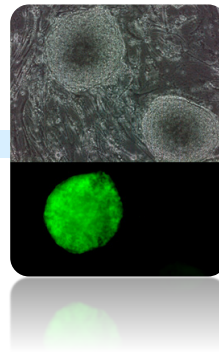
The targeting vector was electroporated in the proprietary BALB/N F122 embryonic stem cell (ESCs) line.

Transfected ES clones were submitted to neomycin selection (G418) and 47 resistant ES clones were isolated. The clones were then submitted to the screening process allowing secured identification of those harbouring the expected recombination events at both ends of targeting vector.

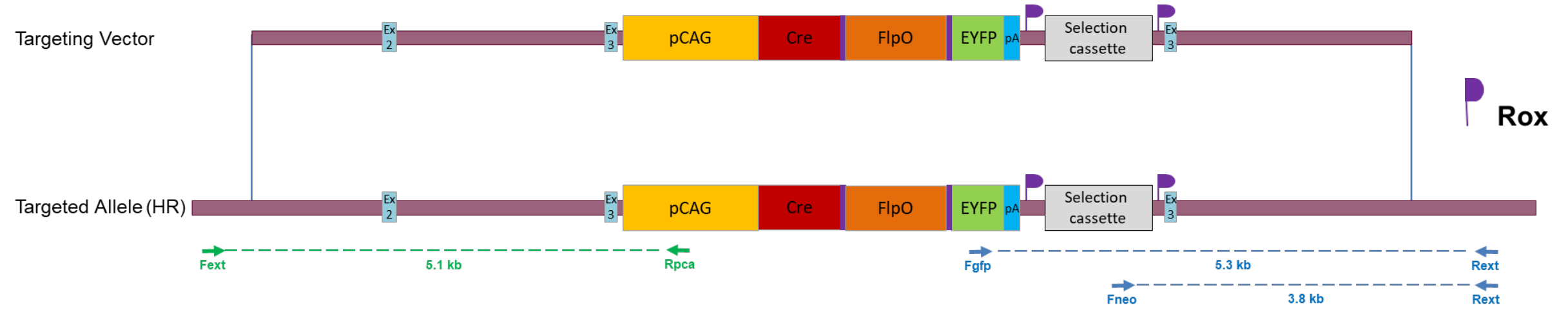
Screening process steps:

1. Identification of candidate recombinant clones by initial 3' Long-Range PCR
2. Two of 3' PCR positive clones were confirmed for 5' recombination event by Long-Range PCR
3. Positive clones in step 2 were further validated by Southern blot analysis using internal and external probes
4. The karyotype of the validated clone was verified using Giemsa staining

# Long range PCR screening – strategy

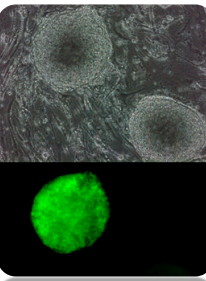


## Schematic 5' and 3' PCR screening strategy

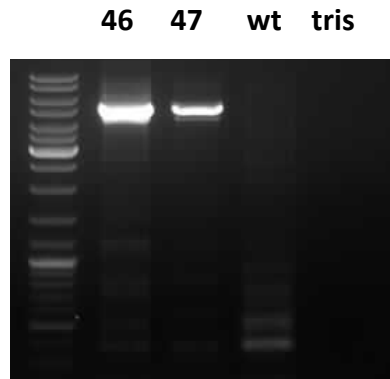
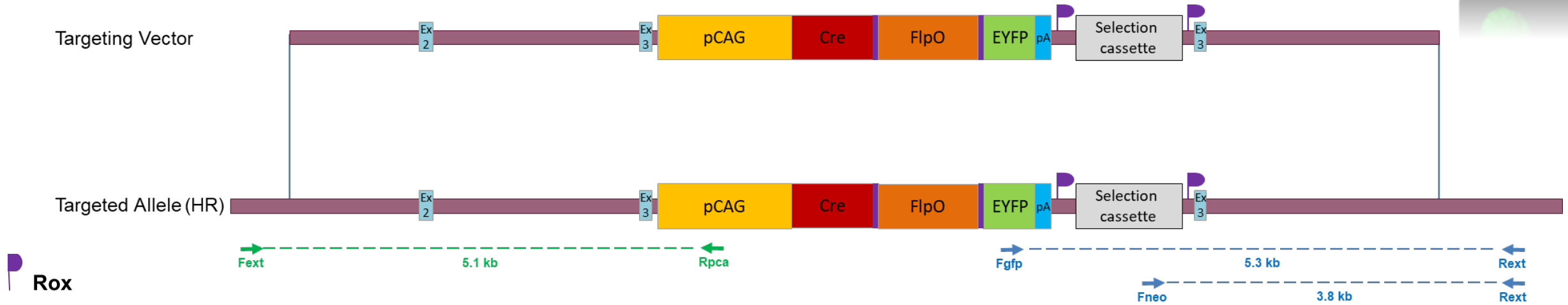


PCR	Primer Name	Primer sequences	PCR product size
5' PCR	Fext	GAGTTTTAGGCCAGTTTAGGATCCA	5.1 kb
	Rpca	GCAGAACGTGGGGCTCACCTCGACC	
3' PCR	Fgfp	CCCGTGCTGCTGCCCGACAACCACT	5.3 kb
	Rext	AATATGGCCGCTCTAAATAGAAACAAGAGCTCAGG	
3' PCR	ADB296	AGGGGCTCGCGCCAGCCGAAGCTGTT	3.8 kb
	Rext	AATATGGCCGCTCTAAATAGAAACAAGAGCTCAGG	

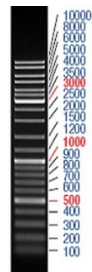
# Recombinant ES validation by Long Range PCR



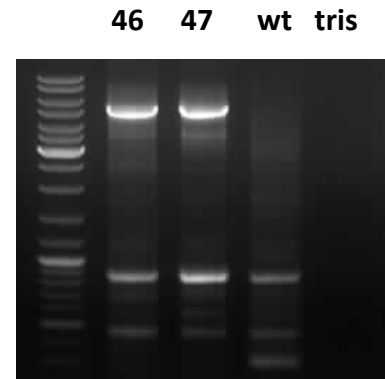
## Confirmation and Validation of candidate recombinant ES clones by 5' and 3' PCRs



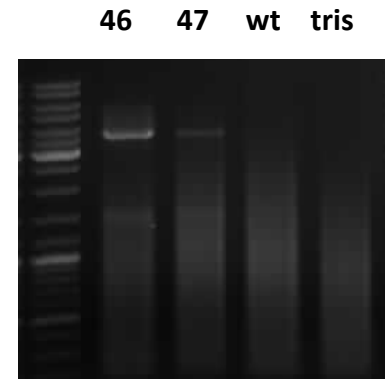
Pcr Fext – Rpca: 5.1 kb



Ladder pattern



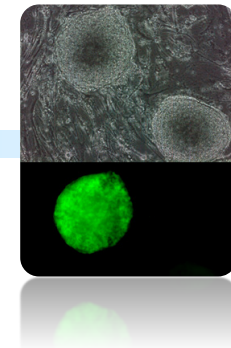
Pcr Fgfp – Rext : 5.3 kb



Pcr Fneo – Rext : 3.8 kb

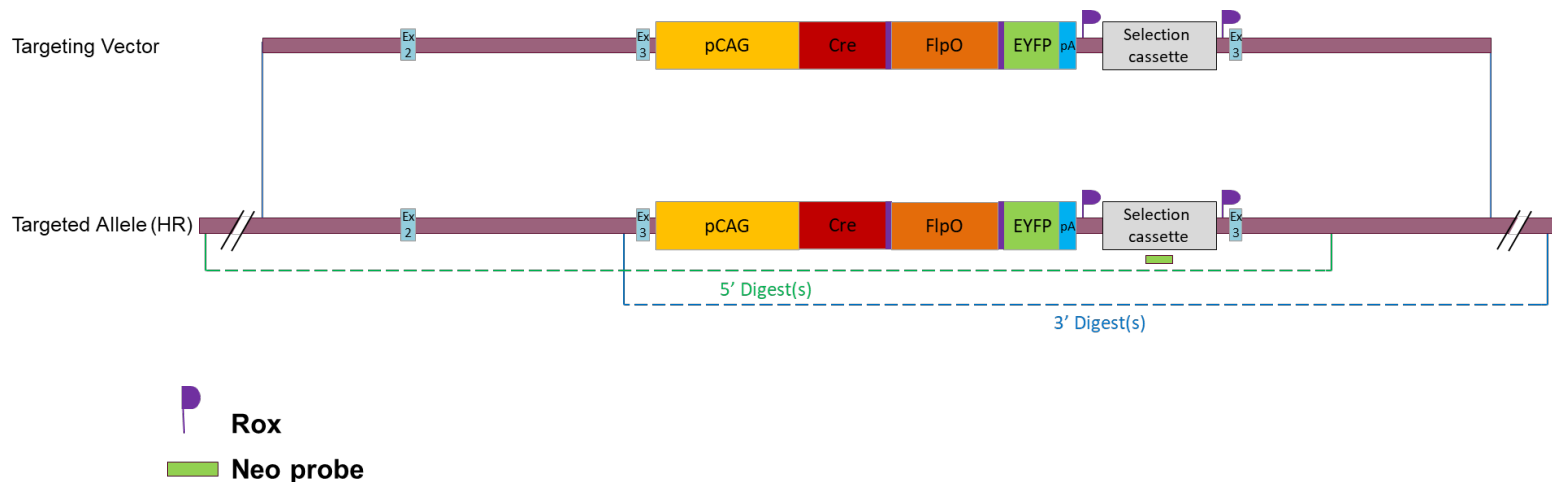
Two candidate clones were analysed by 5' and 3' PCRs screening.  
Two clones (clones #46 and #47) were confirmed.

# Recombinant ES clones validation by Southern Blot – Internal probe



## Schematic Southern Blot validation strategy

Digests on the scheme illustrate the position of the chosen restriction sites relative to the probe. They don't show the exact position of the restriction sites.



## Digestions used to validate the 5' and 3' insertion

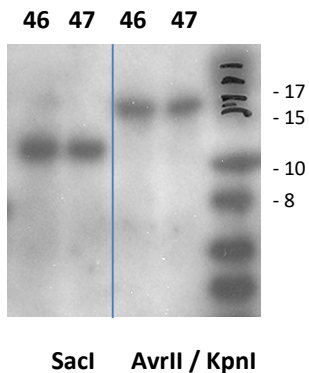
Probe		Genomic DNA digest	Targeted Allele (kb)
Neo	5' digest	SacI	12
		AvrII / KpnI	18
	3' digest	BglII	4.9
		XmnI	8.9

## Neo probe sequence

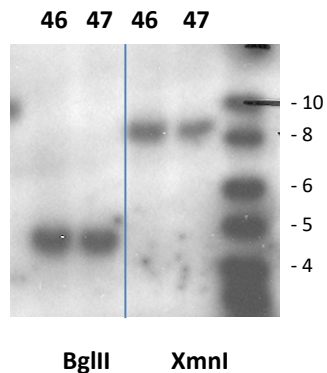
```

ATGATTGAACAAGATGGATTGCACGCAGGTTCTCCGGCCGCTTGGGTGGAGAGGCTATTCGGC
TATGACTGGGCACAACAGACAATCGGCTGCTCTGATGCCGCCGTGTTCCGGCTGTCAGCGCAG
GGGCGCCCGGTTCTTTTTGTCAAGACCGACCTGTCCGGTGCCCTGAATGAACTGCAGGACGAG
GCAGCGCGGCTATCGTGGCTGGCCACGACGGGCGTTCCTTGCGCAGCTGTGCTCGACGTTGTC
ACTGAAGCGGGAAGGGACTGGCTGCTATTGGGCGAAGTGCCGGGGCAGGATCTCCTGTCATCT
CACCTTGCTCCTGCCGAGAAAGTATCCATCATGGCTGATGCAATGCGGCGGCTGCATACGCTT
GATCCGGCTACCTGCCCATTCGACCACCAAGCGAAACATCGCATCGAGCGAGCACGTA CT CGG
ATGGAAGCCGGTCTTGTGCGATCAGGATGATCTGGACGAAGAGCATCAGGGGCTCGCGCCAGCC
GAACTGTTCCGCAAGGCTCAAGGCGCGCATGCCCGACGGCGAGGATCTCGTCGTGACCCATGGC
GATGCCTGCTTGCCGAATATCATGGTGGAAAATGGCCGCTTTTCTGGATTTCATCGACTGTGGC
CGGCTGGGTGTGGCGGACCGCTATCAGGACATAGCGTTGGCTACCCGTGATATTGCTGAAGAG
CTTGCGCGGAATGGGCTGACCGCTTCTCGTGCTTTACGGTATCGCCGCTCCCGATTGCGAG
CGCATCGCCTTCTATCGCCTTCTTGACGAGTCTTC
    
```

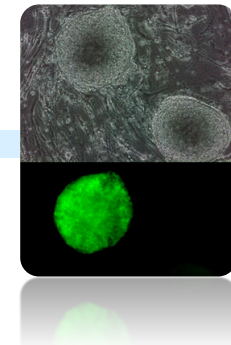
### Southern blot - Neo 5'



### Southern blot - Neo 3'

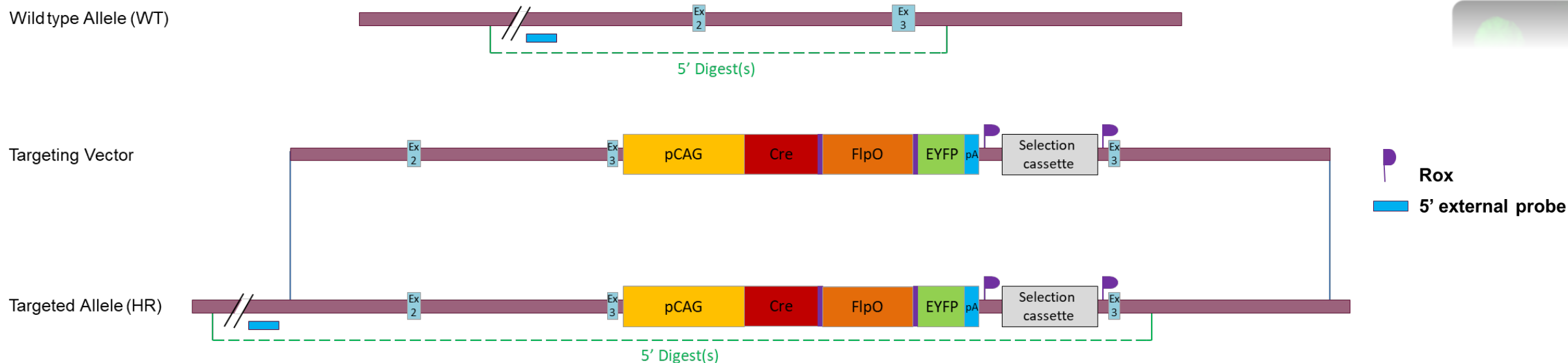


# Recombinant ES clones validation by Southern Blot – External probe

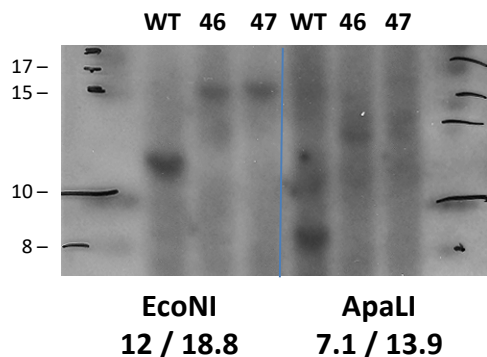


## Schematic Southern Blot validation strategy

Digests on the scheme illustrate the position of the chosen restriction sites relative to the probe. They don't show the exact position of the restriction sites.



## Southern blot – 5' probe



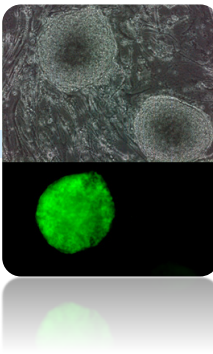
## Digestions used to validate the 5' and 3' insertion

Probe	Name	Genomic DNA digest	WT allele (kb)	Targeted Allele (kb)
5' external probe	5' first digest	EcoNI	12	18.8
	5' second digest	ApaII	7.1	13.9

## 5' PROBE SEQUENCE

```
GGCAGAGTTCACATTGGATTTGTCTTTAATTGAAACAGTCTTTGAAGTTCTTAG
GTAGGTTACCTGTAGAACCTGTAACCAGGCATTGAGCTATTTTAGTAGGTGCAG
TAGTCTGATGAAACCTGGGTGTGATAGGCTTAAGGCAGAAACAGAGTGAGTGGG
CAGAGGGTATGAGAATGCCTCAGGAAAGTCATTGTGGTATTCTAATTTTGATTC
TAAGGTACTTTGAAAATAATCCAGCTTTATTTTTGTCAGGTAGTACATAAGTTCT
CATGAGCCTGGCCTGAATATTTATTGTTGTTATCTTATAAAAATTGAGCAGTCTG
TCATCTTTCCCCAAAACATGAACTAAAATTTGCTTTATAGAACTTACAAAAATG
CTTTTTTTGTAATAACTCTTTGAATATTCCTTTGTATGATACAAATGCC
```

## ■ Aneuploidy screening in ES recombinant clones



Selected recombinant ES cells clones were karyotyped by ddPCR as described in Codner *et al.*<sup>1</sup> and by Giemsa metaphase staining. Results of aneuploidy analysis are presented in the table below.

Clone ID	ddPCR	Giemsa
#46	Pass	Pass
#47	Pass	Pass

<sup>1</sup> Codner, G.F., Lindner, L., Caulder, A., Wattenhofer-Donzé, M., Radage, A., Mertz, A., Eisenmann, B., Mianné, J., Evans, E.P., Beechey, C.V., Fray, M.D., Birling, M.-C., Hérault, Y., Pavlovic, G., Teboul, L  
Aneuploidy screening of embryonic stem cell clones by metaphase karyotyping and droplet digital polymerase chain reaction.  
BMC Cell Biology 2016 doi:10.1186/s12860-016-0108-6

# 5 MICROINJECTION & BREEDING



- Microinjection
- Breeding to F1 generation

## ■ Microinjection



- The ES cells used in the injection experiment were originally derived from a BALB/cN mouse strain (which have white coat colour). These cells were injected into blastocysts derived from an C57BL/6N strain, which have a black coat colour. The resulting offspring are thus chimeras of two different cell types (ES cell-derived cells and host blastocyst-derived cells) and the degree of chimerism was monitored by the percentage of light and dark patches on these animals.
- Recipient blastocysts were isolated from mated C57BL/6N females (Health status SPF Specific Pathogens Free).
- Recombinant ES clones #46 and #47 validated in previous project phase were injected into blastocysts to generate chimeric males. The results are presented in the table below.

Clone ID	Number of chimeric males identified according to chimerism rate (Number of chimeric males bred to F1 generation)			
	5 - 40%	45% - 55%	60-100%	Total
#46	0	1	2	3
#47	0	0	1	1

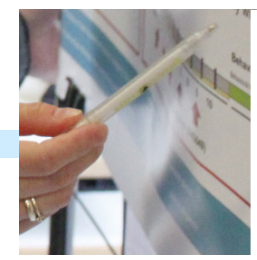


## ■ Breeding to F1 generation



- Three highly chimeric males generated in the previous phase by blastocyst injection of the ES clones were mated with wild-type BALB/cN females (health status SPF – Specific Pathogen Free) to investigate whether the recombined ES cells have contributed to the germ layer.
- Germ line transmission was obtained the: 25/12/2012
- Allele nomenclature (following MGI guidelines) : **BALB/cN Hprt<sup>tm1(pCAG-Cre-Flpo-EYFP)</sup>Ics**

# 6 SEQUENCE OF THE DELIVERED ALLELE



AGATGTTATAGTGTACTCTCCTCTCCCTCCCCTCCCCTCCTCCTCAGCAGGATCTTGTATGTAGCCCTAAGGGATCTAATACTTACTCTGTAGTCCAGGCTGTCTTACAGTCATGAGACACACCATCATGCTTGGCATGTTTCTGTCTGAAAGGTATCTGGTTTTAATTTTGGTT  
GAGCTGGTGTGTTGGTGTATATCTGTAATCTCAGCACAGGCAAGTAGAGACAAGGATTATGATTACAGTCCAGCCTGGGCTATTTAGAGGGATTCTTTAATCTTTCTCATGCCCAAATCTTACCTTTGGTATATGAAAAATAGTCTCCACTTCTGCAAAATATTGCTTTATGAAG  
TAAGAATTCCCTTCATAGAGACAAGGAATGTGTCTGTAAAAGTTAATGTGTAAGAAGTATTTGTTATAAAAGATAAATATTAGAATCTTCTTTTTAATTCCTGATTTTATTTCTATAGGACTGAAAGACTTGCTCGTCGACATTGATTATTGACTAGTTAATAAGTAAATCAATTACG  
GGGTCAATTAGTTCATAGCCCATATATGGAGTTCGCGTACATAACTACGGTAAATGGCCCGCTGGCTGACCGCCCAACGACCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAA  
CTGCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCTATTGACGTCAATGACGGTAAATGGCCCGCTGGCATTATGCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGGTGAGGTTGAGCCCCAC  
GTTCTGCTTCACTCTCCCATCTCCCCCTCCCACCCCAATTTTGTATTTATTTATTTTAAATTTTGTGCAGCGATGGGGGCGGGGGGGGGGGGGCGCGCCAGGGCGGGGCGGGGCGAGGGGCGGGGCGGGGCGAGGCGGAGAGGTGCGGCGGCAGCCA  
ATCAGAGCGGCGCTCCGAAAGTTTCTTTATGGCAGGGCGGCGGCGGCGCCCTATAAAAAGCGAAGCGCGCGGGCGGGAGTCTGCTGCTTGCCTTCCGCCGTGCCCGCTCCGCGCCCTCGCGCCGCCCGCCCGCTGACTGACCGGTTACTCCACA  
GGTGTAGCGGGCGGGACGGCCCTTCTCCTCGGGCTGTAATTAGCGCTTGGTTAATGACGGCTCGTTTCTTTCTGTGGCTGCGTGAAAGCCTAAAGGGCTCCGGGAGGGCCCTTTGTGCGGGGGGAGCGGCTCGGGGGTGCCTGCTGTGTGTGTGCTGGGGAGCGCG  
CGTGTGCGCCCGCGCTGCCGCGGCTGTGAGCGTGTGCGGCGCGGCGGGGCTTTGTGCGCTCCGCTGTGCGGAGGGGAGCGCGCCGGGGCGGTGCCCGCGTGTGCGGGGGGCTGCGAGGGGAACAAAGGCTGCTGCGGGGTGTGTGCTGGGGGGTGTGCA  
GGGGGTGTGGGCGCGGCGGTGCGGGCTGTAACCCCTTGCACCCCTCCCGAGTTGCTGAGCACGCGCCGCTTCCGGTGTGCGGGGCTCCGTGCGGGGCTGCGCGGGGCTGCGCGGGGCGGGGGTGGCGGAGGTGGGGGTGCCGGCGGGGCGGGGCGCC  
TCGGGCCGGGGAGGGCTCGGGGAGGGGCGCGCGGCCCGGAGCGCGGCGGCTGTGAGGCGCGGCGAGCCGAGCCATTGCTTTTATGTAATCGTGTGAGAGGGGCGAGGGACTTCTTTGTCCAAATCTGGCGGAGCCGAAATCTGGGAGGCGCGCCGCCACCC  
TCTAGCGGGCGCGGCGAAGCGGTGCGGCGCCGGCAGGAAGGAAATGGGCGGGGAGGGCTTCTGTCGTCGCGCGCCGCGCTCCCTTCTCATCTCCAGCCTCGGGGCTGCCGACGGGGACGGCTGCCTTCCGGGGGGACGGGGCAGGGCGGGGTTCCGGCTTCTGGCGT  
GTGACCGGGGCTCTAGAGCTCTGCTAACCATGTTTCATGCTTCTTTTCTACAGCTCTGGCAACGTGCTGGTTATTGTGTGTCTCATATTTGGCAAAGAAATTCACACCATGTCCAATTTACTGACCGTACACCAAAATTTGCCTGCATTACCGTGCATGCAACGAGTG  
ATGAGGTTTCGCAAGAACCTGATGGACATGTTACGGGATCGCCAGGCGTTTTCTGAGCATACTGGAAAATGCTTCTGTCCGTTTCCCGTGTGGGCGGCATGGTGAAGTTGAATAACCGGAAATGGTTTCCCGCAGAACCTGAAGATGTTCCGATTATCTTATATCTCAGG  
CGCGGTCTGGCAGTAAAAATATCCAGCAACATTTGGGCCAGCTAAACATGCTTTCATCGTCCGCTCCGGCTGCCACGACCAAGTGACAGCAATGCTGTTTCACTGGTTATGCGGCGGATCCGAAAAGAAAACGTTGATGCCGGTGAACGTGCAAAACAGGCTTAGCGTTCGAAC  
CACTGATTTCCAGCAGGTTCTTCACTCATGGAAAATAGCGATCGTCCAGGATATACGTAATCTGGCATTCTGGGATTGCTTATAACACCCTGTACGTATAGCCGAAATGCCAGGATCAGGGTTAAAGATATCTACGTACTGACGGTGGGAGAAATGTTAATCCATATTGG  
CAGAACGAAAACGCTGGTTAGCACCGCAGGTGTAGAGAAGGCATTAAGCTGGGGTAACTAACTGGTGTGAGCGATGGATTTCCGCTCTGTGTGTAGCTGATGATCCGAATAACTACTGTTTTGCCGGGTGAGAAAAATGGTGTGCCGCGCATCTGCCACCAGCCAGCTAT  
CACTCGCGCCCTGGAAGGGATTTTGAAGCAACTCATCGATTGATTACGGCGCTAAGGATGACTCTGGTGTGAGAGATACCTGGCCTGGTGTGACACAGTCCCGTGTGCGAGCCGCGCAGATATGGCCCGCTGGAGTTTCAATACCGGAGATCATGCAAGCTGGTGGCTGG  
ACCAATGTAATATTGTATGAATATATCCGTAACCTGGATGTGAAACAGGGGCAATGGTGTGCGCTGCTGGAAGATGGGATCAGCTGTGAATTTGACCTTCTAAGCTTGTGGGAGACGTGAGTCCAACCTGGGCCATGAGCCAGTTCGACATCTGTGCAAGACCC  
CCAAAGGTGCTGGTGTGCGCAGTTCTGTGGAGAGATTGAGAGGCCAGCGCGGAGAAAGATGCCAGCTGTGCGCCGAGCTGACCTACCTGTGCTGGATGATACCCCAACGGCACCCCATCAAGAGGGCCACCTTATGAGCTACAACACCATCATCAGCAACAGCCTGAGCT  
TCGACATCGTGAACAAGACCTGCGATTCAAGTACAAGACCCAGAAAGGCTGCTGAGGCGACCTGGAAGGAGCTGATCCCGCTGGGAGTTCCACCATCTTACAACGCGCAGAACAGCAGGACATCACCAGCATCGTGTCCAGCTGACCTGAGCTGAGTTGCGAG  
AGCAGCGAGGAGGCCGACAAGGGCAACAGCCACAGCAAGAAGATGCTGAAAGCCCTGCTGTCCGAGGGCGAGAGCATCTGGGAGATCACCAGAAAGATCTGAAACAGCTTCTGAGTACACCAGCAGGTTCAACAAGACCAAGCCCTGTACCAAGTCTGTTCTGCCACATTC  
TCAACTGCGCGAGGTTCAAGCAGATCAAGAACCTGGACCCCAAGAGCTTCAAGCTGGTGCAGAACAAGTACCTGGGCGTGTATTTCAAGTGCCTGGTACCAGAGACCAAGACAAGCGTGTCCAGGCACATCTACTTTTTCAGCGCCAGAGGACGAGTCCAGCCCTGGTGTACCTG  
GACGAGTCTGAGGAACAGCGAGCCGCTGCTGAAGAGAGTGAACAGGACCAGCAACAGCAGCAGCAACAAGCAGGAGTACCAGTGTGAAGGACAACCTGGTGTGCGAGCTACAACAAGGCCCTGAAGAAGAACGCCCCCTACCCATCTTCCGATCAAGAAGCGCCCTAAG  
AGCCACATCGGCAGGCACCTGATGACCAGCTTTCTGAGCATGAAGGGCCTGACCGAGCTGACAAACGTGGTGGGCAACTGGAGCGACAAGAGGGCCTCCGCGTGGCCAGGACCACCTACACCACAGATCACCGCCATCCCGACCACTACTTCCGCCCTGGTGTCCAGTACTA  
CGCCTACGACCCCATCAGCAAGGAGATGATCGCCTGAAGGACGAGACCAACCCATCGAGGAGTGGCAGCAGATCGAGCAGCTGAAGGGCAGCGCGAGGGCAGCATCAGATACCCCGCTGGAACGGCATCATCAGCCAGGAGGTGCTGGACTACTGAGCAGCTACATCAA  
CAGGGGATCGAGGGCAGAGGAAGTCTTAAACATGCGGTGACGTGGAGGAGAATCCCGCCCTATGGTGTGAGCAAGGGCGAGGAGCTTACCAGGGGTGGTGGCCATCTGCTGAGCTGGACGGCGACTTAAACGGCCACAAGTTACAGCGTGTCCGGCAGGGCGAGGGC  
GATGCCACCTACGGCAAGCTGACCCTGAAGTTCATCTGCACCACCGCAAGCTGCCGTGCCCTGGCCACCCTCGTGACCACCTTGGCTACGGCCTGCAGTGTCTCGCCGCTACCCGACCATGAAGCAGCAGACTTCTTCAAGTCCGCCATGCCGAAGGCTACGTCCAG  
AGCGCACCATCTTCAAGGACGACGGCAACTACAAGACCCGCGCGGAGGTGAAGTTCGAGGGCGACACCCTGGTGAACCGCATCGAGCTGAAGGGCATCGACTTCAAGGAGGACGGCAACATCTGGGGCACAAGCTGGAGTACAACAGCCACAACGCTTATATCAT  
GGCCGACAAGCAGAAGAACGGCATCAAGGTGAATTCAGATCCGCCACAACATCGAGGACGCGAGCTGCAGTGTCCGACCACTACCAGCAGAACACCCCATCGGCGACGGCCCGTGTGTGCTGCCGACAACCACTACTGAGTACCAGTCCGCGCTGAGCAAAGACCC  
AACGAGAAGCGCATCACATGGTCTGTGGAGTTCGTGACCGCCCGGGATCACTCTGGCATGGACGAGCTGTACAAGTAAGGATCCAGATCTTATAAAGCAGAATTGTTTATTGAGCTTATAATGGTTACAATAAAGCAATAGCATCACAATTTACAATAAAGCATT  
TTTTCACTGCATTAGTTGTGGTTGTCCAACCTCATCAATGTATCTTATCATGTCTGGTGTGAGGCGGGCCTAACTTAAATAATTGGCATTATTTAAAGTTAGCCGGCCGAGATGTCATGAAGGAGATGGGAGGCCATCATTGTGGCCCTGTGTGCTCAAGGGGGGCTAT  
AAGTTCTTTGCTGACCTGCTGGATTACATAAAGCACTGAATAGAAAATAGTATAGATCCATTCTATGACTGTAGATTTTATCAGACTGAAGAGCTACTGTGTAAGTATAATTAACCTATAATTAATAAAGGACCTTCAAGTTTATCTATATTTTTTAACTGTGCAAACTAT  
GCTACATAATCAACTTACTTATTGATTAATATATAGTAATAGTTGGAATTAATATTACATATTTATTTTTCTGTGCTGGGGTAGAACCCAGGGAACTAAGTACTGTAATAACCGAGCTTATTATCTAGTATTTTTTAAATTTTTTTTTTTTTTTTTTTTGGTTTTTTCGAGACAAGGTTT  
CTCTGTGTAGCCCTGGCTGTCTGGAACCTACTCTGTAGACCAGGCTGGCTCGAACCTCAGAAATCTGCC

pCAG

Cre

2A

FlpO

EYPF

Rox

Sv40 polyA



## REPORT REDACTION & VALIDATION

Prepared by Romain LORENTZ, IE

Verified by Marie-Christine BIRLING, PhD

## CONTACT US

By email at [mutagenesis@igbmc.fr](mailto:mutagenesis@igbmc.fr)

By phone at +33 (0)3 88 65 56 57

[www.phenomin.fr](http://www.phenomin.fr)