





Allele Description

This is a CRISPR/Cas9 induced mutation creating a conditional knock-out by floxing critical exon, ENSMUSE00000745494 of *Myocd*. The stock was generated at MRC Harwell via pronuclear injection of CRISPR/Cas9 reagents into 1-cell stage embryos.

<u>qPCR Copy Counting Genotyping Strategy</u>

The genotyping strategy presented here has been optimized for reagents and conditions used by the Genotyping Core at MRC Harwell. To genotype animals, we recommend researchers validate the assay independently. PCR cycling temperature and times may require additional optimization based on the specific genotyping reagents used.

Samples are genotyped using qPCR copy counting with both a wildtype loss of allele (WT-LOA) and a mutant assay against a known reference assay (*Dot1l* on chromosome 10; 2 copies present). Samples for this line are genotyped using the following primers and probe:

- Wildtype assay where the probe is situated within the deleted region/at the LoxP insertion site.
- Mutant assay where the probe binds to the inserted LoxP sequence

For autosomal genes that have been targeted, the following results would be expected:

Genotype of the Modified allele	WT Assay	Mutant Assay
Wildtype	2	0
Heterozygous	1	1
Homozygous mutant	0	2





Myocd-Flox-3'-WT1 assay (FAM labelled)

TGCCCCCCCCAGAACTAGGGAAAGGTACCCTGAACTCTCATGCCAAGCTGGACCAATTCT<u>GACT</u> <u>TCTCTTTTGTCTTTTACCTCTC</u>CTTCAATTTCCCAGTTGCCTGTTTCT <u>ACTTAGAGGCTGGCTTTGAAT</u>TTGTTTTTATATCCAGTAGGGAAGAAATTCCAAAGTATTTCATGTT

Probe sequence is in bold and shaded grey Primer sequences are in bold and underlined

Oligo Myocd-				
Flox	5' label	Sequence 5' → 3'	3' label	Oligo Type
Myocd-Flox-				
WT_F	n/a	GACTTCTCTTTTGTCTTTTACCTCTC	n/a	WT Forward
Myocd-Flox-			ZEN-	
WT_PROBE	FAM	CTTCAATTTCCCAGTTGCCTGTTTCT	IBFQ	WT Probe
Myocd-Flox-				
WT_R	n/a	ATTCAAAGCCAGCCTCTAAGTA	n/a	WT Reverse

Myocd-Flox-3'-MUT2 assay (FAM labelled)

Lower case letters denote the inserted LoxP sequence Probe sequence is in bold and shaded grey Primer sequences are in bold and underlined

Oligo Myocd-				
Flox	5' label	Sequence 5' \rightarrow 3'	3' label	Oligo Type
Myocd-Flox-				
MUT_F	n/a	GCCAAGCTGGACCAATTCTG	n/a	Mutant Forward
Myocd-Flox-				
MUT_PROBE	FAM	AAGTTATCGCCGGCGGGTCTGA	BHQ1	Mutant Probe
Myocd-Flox-				
MUT_R	n/a	CAAAGCCAGCCTCTAAGTAGAC	n/a	Mutant Reverse





Dot1l internal control (VIC labelled)

CTGATGGGTGTGGGCAGATCCTACAGAGTCCCATTGGCCACCATGTGTGCTACGCCTGAAATAAAGCCTTGCC **CCAGCACGACCATT**CAGGGCCAGCTCTCAAGTCGACTGTAAGATGAAGCATAAGGATGCCAACTACTAACA GAAAACGACTAGAGGGGAAAAGAACAAGGAAACAGAAGACGCAGCACTCCGGCTTCCCTGGGTTGGCCAGT CACCCTATGA

Oligo Myocd-				
Flox	5' label	Sequence $5' \rightarrow 3'$	3' label	Oligo Type
Dot1l_Forward	n/a	GCCCCAGCACGACCATT	n/a	WT Forward
Dot1l_Probe	VIC	CCAGCTCTCAAGTCG	BHQ	WT Probe
Dot1l_Reverse	n/a	TAGTTGGCATCCTTATGCTTCATC	n/a	WT Reverse

Probe sequence is in bold and shaded grey Primer sequences are in bold and underlined

DNA extraction method

DNA is extracted from ear clips using Applied Biosystems Taqman Sample-to-SNP Kit and qPCR run using 1:10 dilution from the crude preparation.

q <u>PCR master mix</u>	1X
Applied Biosystems GTX Taqman master mix	5 µl
Dot1l_Forward (20 μM)	0.225 μl
Dot1l_Reverse (20 μM)	0.225 μl
Dot1l_Probe (5 μM)	0.2 μl
FAM Assay (probe 5 μM & primers 15 μM each)	0.3 μl
ddH20	1.55 μl
DNA (1:10 dilution of ABI Sample-to-SNP prep)	2.5 μl

Each sample is ran in technical duplicate. Seven WT and/or mutant controls are also included in duplicate along with non-template controls.

qPCR cycling conditions

qPCR instrument: Applied Biosystems 7500/7900 or ThermoFisher QuantStudio 7

95°C for 20 sec Then 40 cycles of; 95°C for 3 sec 60°C for 30 sec





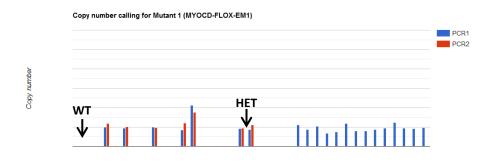


<u>Analysis</u>

Copy number

The results are analysed using CopyCaller software v2.0 from Applied Biosystems or inhouse software that is based on CopyCaller v2.0.

<u>Myocd-Flox'-WT1 and Myocd-Flox-MUT1 assays copy called results, image showing copy</u> <u>number chart for WT and Mutant assays (Task 306474 results)</u>



Copy number calling for WT LOA 1 (MYOCD-FLOX-EM1)

Version No.	1
Date:	23/06/2020
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Approved by:	Adam Caulder 01/10/20