ACAN-NTVEGA-EM1+EM2

Allele Description

This is a CRISPR/Cas9 induced mutation for ACAN-NTVEGA created to introduce the E392A change together with a silent change to prevent re-processing of the engineered allele. The stock was generated at MRC Harwell via microinjection of CRISPR/Cas9 reagents into 1-cell stage embryos.

qPCR Copy Counting Genotyping Strategy

The genotyping strategy presented here has been optimized for reagents and conditions used by the Genotyping Core at MRC Harwell. To genotype animals, we recommend researchers validate the assay independently. PCR cycling temperature and times may require additional optimization based on the specific genotyping reagents used.

Samples are genotyped using qPCR copy counting with both a wild type and a mutant assay against a known reference assay (*Dot1l* on chromosome 10; 2 copies present). Samples for this line are genotyped using the following primers and probe:

- Wild type (WT) assay with probe and reverse primer binding to the WT bases mutated in the mutant allele.
- Mutant assay with probe and reverse primer binding to the G601R, F606Y and R609H point mutations.

For autosomal genes that have been targeted, the following results would be expected:

Genotype of the Modified allele	WT Assay	Mutant Assay
Wildtype	2	0
Heterozygous	1	1
Homozygous mutant	0	2

ACAN-NVTEGA-WT1 assay (FAM labelled)

GAAGGAAATGAGGCACATTCAGCTCTTGTCCCCTCAAAACAGCCCTGAGCTCAGCACTAAAGCCTGT
CATTGCCTCCTGTCCCAGGTGAAGATTTTGTAGACATCCCAGAAAACTTCTTCGGGGTGGGTGGTGA
AGACGACATCACCATCCAGACGGTGACCTGGCCAGACCTGGAGCTGCCCCTGCCCCGTAATG**TCAC
AGAGGGAGaGGCc**CTGGGCAG**CGTGATCCTTACAGCAAAGCCCATCT**TTGACCTGTCCCCCACTAT
CTCAGAGCCGGGGGAGGCCCTCACACTTGCCCCTGAAGTGGG**GAGCACAGCCTTCCCAG**AGGCTG
AGGAGAGAACTGGAGAAGCCACCAGACCCTGGGGCTTTCCTGCAGAAGTCACACGTGGGCCGGAC
TCTGCCACTGCCTTCGCCAGTGAGGACCTGGTAGTGCGAGTGACCATCTCTCCAGGTGCAGCTGAAG

Lower case letters denote bases changed in the mutant allele.
Probe sequence is in bold and shaded grey.
Primer sequences are in bold and underlined.

Oligo Name	5' label	Sequence 5' → 3'	3' label	Oligo Type
ACAN-NVTEGA-WT_F	n/a	**TCACAGAGGGAGAGGCC**	n/a	Wild type Forward
ACAN-NVTEGA-WT_PROBE	FAM	**CGTGATCCTTACAGCAAAGCCCATCT**	ZEN/IBFQ	Wild type Probe
ACAN-NVTEGA-WT_R	n/a	**CTGGGAAGGCTGTGCTC**	n/a	Wild type Reverse

ACAN-NVTEGA-MUT1 assay (FAM labelled)

GAAGGAAATGAGGCACATTCAGCTCTTGTCCCCTCAAAACAGCCCTGAGCTCAGCACTAAAGCCTGT
CATTGCCTCCTGTCCCAGGTGAAGATTTTGTAGACATCCCAGAAAACTTCTTCGGGGTGGGTGGTGA
AGACGACATCACCATCCAGACGGTGACCTGGCCAGACCTGGAGCTGCCCCTGCCCCGTAATG**TCAC
AGAGGGAGcGGCt**CTGGGCAG**CGTGATCCTTACAGCAAAGCCCATCT**TTGACCTGTCCC**CCACTATCTC
AGAGCCGGG**GGAGGCCCTCACACTTGCCCCTGAAGTGGGGAGCACAGCCTTCCCAGAGGCTGAGG
AGAGAACTGGAGAAGCCACCAGACCCTGGGGCTTTCCTGCAGAAGTCACACGTGGGCCGGACTCTG
CCACTGCCTTCGCCAGTGAGGACCTGGTAGTGCGAGTGACCATCTCTCCAGGTGCAGCTGAAG

Lower case letters denote bases changed in the mutant allele.
Probe sequence is in bold and shaded grey.
Primer sequences are in bold and underlined.

Oligo Name	5' label	Sequence 5' → 3'	3' label	Oligo Type
ACAN-NVTEGA -MUT_F	n/a	**TCACAGAGGGAGCGGCT**	n/a	Mutant Forward
ACAN-NVTEGA -MUT_PROBE	FAM	**CGTGATCCTTACAGCAAAGCCCATCT**	BHQ	Mutant Probe
ACAN-NVTEGA -MUT_R	n/a	**CCCGGCTCTGAGATAGTGG**	n/a	Mutant Reverse

Dot1l internal control (VIC labelled)

CTGATGGGTGTGGGCAGATCCTACAGAGTCCCATTGGCCACCATGTGTGCTACGCCTGAAATAAAGCCTT**GCC CCAGCACGACCATT**CAGGG**CCAGCTCTCAAGTCG**ACTGTAA**GATGAAGCATAAGGATGCCAACTA**CTAACA GAAAACGACTAGAGGGGAAAAGAACAAGGAAACAGAAGACGCAGCACTCCGGCTTCCCTGGGTTGGCCAGT CACCCTATGA

Oligo Name	5' label	Sequence 5' → 3'	3' label	Oligo Type
Dot1l_Forward	n/a	**GCCCCAGCACGACCATT**	n/a	WT Forward
Dot1l_Probe	VIC	**CCAGCTCTCAAGTCG**	BHQ	WT Probe
Dot1l_Reverse	n/a	**TAGTTGGCATCCTTATGCTTCATC**	n/a	WT Reverse

Probe sequence is in bold and shaded grey
Primer sequences are in bold and underlined

DNA extraction method

DNA is extracted from ear clips using Applied Biosystems Taqman Sample-to-SNP Kit and qPCR run using 1:10 dilution from the crude preparation.

qPCR master mix   1X

Applied Biosystems GTX Taqman master mix	5 µl
Dot1l_Forward (20 µM)	0.225 µl
Dot1l_Reverse (20 µM)	0.225 µl
Dot1l_Probe (5 µM)	0.2 µl
FAM Assay (probe 5 µM & primers 15 µM each)	0.3 µl
ddH20	1.55 µl
DNA (1:10 dilution of ABI Sample-to-SNP prep)	2.5 µl

Each sample is ran in technical duplicate. Seven WT and/or mutant controls are also included in duplicate along with non-template controls.

qPCR cycling conditions

qPCR instrument: Applied Biosystems 7500/7900 or ThermoFisher QuantStudio 7.

95°C for 20 sec
Then 40 cycles of;
95°C for 3 sec
60°C for 30 sec

Analysis

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The results are analysed using CopyCaller software v2.0 from Applied Biosystems or in-house software that is based on CopyCaller v2.0.

No example available yet

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