

**Name of Mouse model or mutation:****CSF1R-I792T-EM1-B6****Description:**

Point mutation model made using CRISPR/Cas9.

**Type of mutation:**

SNP: I792T

**Sequence details****WT**

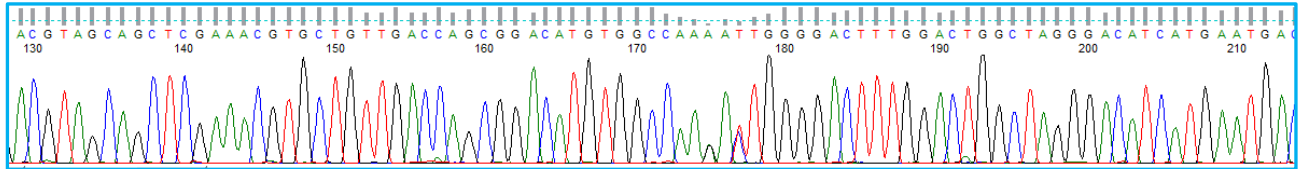
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CGGGACGTAGCAGCTCGAAACGTGCTGTTGACCAGCGGACATGTGGCCAAGATTGGGGACTTTGG  
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TGCTAAGCCTGTTTTAGGTGCAGCCTAGGGCTGACCCGTCTTTGGCTATGCCATCGGTGTCTTGAAC  
CTCATGGACGAAATCTACTCAAAGATGTCGGTGTCCAAGACAGAGCAAGGGCTGGCAGGAGCAATG  
GGATGGTGAGGGCTCTTGAACCG

**Mutant**

AGGGGTCTAACGGGTTGTTGTGTATGCAAATGCTTGGGAAAGCACCTGGTATTGTGTACTTGGAGG  
TGGCAGCTGTTGGTGTGTATGAGGTGGGCCACAGGCTGCTCACAGCACAGGGACTCATTGCCT  
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GAGTAGACAGTAGACTACCAAAACCTGCATCTACTTCAACAGAGACCCAAGACCTCCTGCTCCTCCTC  
TGGTCCTCAGGCCTCAGGGAAGGATAAACTGACTAATAATCTCTCTGCGCTTTCTTCAGTGCATCCAC  
CGGGACGTAGCAGCTCGAAACGTGCTGTTGACCAGCGGACATGTGGCCAA**ACT**GGGGACTTTGG  
ACTGGCTAGGGACATCATGAATGACTCCAACCTATGTTGTCAAGGGCAATGTGAGTGCCGAGAGAGA  
GAGAGAGAGAGGGAGGGA  
GAGAGAGAGAGAGAGAGAGAGAGATTGAGATTGGTTGGGCAGGCTGTGGAGAGCCCTTGACTGACAT

GGTACTGTCTTGT CAGGCCCGCCTGCCTGTAAAGTGGATGGCCCCAGAGAGCATCTTTGACTGCGTC  
TACACAGTTCAGAGTGATGTGTGGTCCTACGGCATCCTCCTCTGGGAGATCTTCTCGCTTGGTGAGC  
TGCTAAGCCTGTTTT CAGGTGCAGCCTAGGGCTGACCCGTCTTTGGCTATGCCATCGGTGTCTTGAAC  
CTCATGGACGAAATCTACTCAAAGATGTCGGTGTCCAAGACAGAGCAAGGGCTGGCAGGAGCAATG  
GGATGGTGAGGGCTCTTGAACCAG

**CSF1R-I792T-EM1-B6 Heterozygous F1 animal sequence trace:**





### QC strategy employed at Harwell to check the edited allele:

Genomic DNA was extracted from ear clip biopsies and amplified in a PCR reaction using the following conditions/primer sequences:

|                             |  |
|-----------------------------|--|
| Geno_Csf1r_I792T_F2 (5'-3') | AGGGGTCTAACGGGTTGTTG   |
| Geno_Csf1r_I792T_R2 (5'-3') | CTGGTTCAAGAGCCCTCACC   |
| Taq Polymerase used         | ThermoFisher SuperFi Taq   |
| Annealing Temperature (°C)  | 64   |
| Elongation time (min)       | 0.5  |
| WT product size (bp)        | 951  |
| Mutant product size (bp)    | 951  |
| Notes                       | Sequence with primers Geno_Csf1r_I792T_F1 (5'-3'; TCAGGCCTGCACAGGTTTAG) and Geno_Csf1r_I792T_R3 (5'-3'; CACTCACATTGCCCTTGACAACATA) |

All amplicons were sent for Sanger sequencing to check for integration of the donor oligo sequence at the target site. F1 sequences should be heterozygous unless on sex chromosome.

### Copy counting by ddPCR

Copy counting of the donor sequence was carried out by ddPCR at the F1 stage to confirm donor oligos were inserted once on target into the genome. The following Taqman assay was used to copy count the donor sequence compared against a VIC-labelled reference assay for Dot1l:

|                        |                        |
|------------------------|------------------------|
| Assay name             | CSF1R-I792T-UNI1       |
| Forward Primer (5'-3') | GCGCTTTCTTCAGTGCATCC   |
| Reverse Primer (5'-3') | TCCCTAGCCAGTCCAAAGTC   |
| Probe (5'-3')          | CCGGGACGTAGCAGCTCGAAAC |
| Label                  | FAM-BHQ1               |

The ddPCR assay recognises both the WT and the mutant allele. WT controls are expected to call at 2 copies and a correct mutation is expected to call at 2 copies for F1 (HET) animals.

|                        |                           |
|------------------------|---------------------------|
| Assay name             | CSF1R-I792T-MUT1          |
| Forward Primer (5'-3') | AGCGGACATGTGGCCAAAAC      |
| Reverse Primer (5'-3') | GCCCTTGACAACATAGTTGGA     |
| Probe (5'-3')          | TTTGGACTGGCTAGGGACATCATGA |
| Label                  | FAM-BHQ1                  |

The ddPCR assay is specific to the mutant I792T allele of the gene. WT controls are expected to call at 0 copies and a correct mutation is expected to call at 1 copy for F1 (HET) animals.

|                        |                           |
|------------------------|---------------------------|
| Reference Assay Name   | Dot1l                     |
| Forward primer (5'-3') | GCCCCAGCACGACCATT         |
| Reverse primer (5'-3') | TAGTTGGCATCCTTATGCTTCATC  |
| Probe (5'-3')          | CCCAACAGGCCTGGATTCTCAATGC |
| Label                  | VIC                       |

VIC-labelled reference assay for Dot1l gene.



## Allele Description

This is a CRISPR/Cas9 induced mutation creating a point mutations; I792T in CSF1R. The stock was generated at MRC Harwell via microinjection of CRISPR/Cas9 reagents into 1-cell stage embryos.

## qPCR Copy Counting Genotyping Strategy

The genotyping strategy presented here has been optimized for reagents and conditions used by the Genotyping Core at MRC Harwell. To genotype animals, we recommend researchers validate the assay independently. PCR cycling temperature and times may require additional optimization based on the specific genotyping reagents used.

Samples are genotyped using qPCR copy counting with both a wild type and a mutant assay against a known reference assay (*Dot1l* on chromosome 10; 2 copies present). Samples for this line are genotyped using the following primers and probe:

- Wild type (WT) assay with probe and reverse primer binding to the WT bases mutated in the mutant allele.
- Mutant assay with probe and reverse primer binding to the G601R, F606Y and R609H point mutations.

For autosomal genes that have been targeted, the following results would be expected:

| Genotype of the Modified allele | WT Assay | Mutant Assay |
|---------------------------------|----------|--------------|
| Wildtype                        | 2        | 0            |
| Heterozygous                    | 1        | 1            |
| Homozygous mutant               | 0        | 2            |



## CSF1R-I792T-WT1 assay (FAM labelled)

TGGTCCTCAGGCCTCAGGGAAGGATAAACTGACTAATAATCTCTCT**GCGCTTTCTTCAGTGCATCCAC**  
**CGGGACGTAGCAGCTCGAAAC**GTGCTGTTGACCAGCGGACATGTGGCCAA**gAtTGGGGACTTTGG**  
**ACTGG**CTAGGGACATCATGAATGACTCCA**ACTATGTTGTCAAGGGCAATGTGAGTGCCGAGAGAGA**

Lower case letters denote bases changed in the mutant allele.  
 Probe sequence is in bold and shaded grey.  
 Primer sequences are in bold and underlined.

| Oligo CSF1R-I792T     | 5' label | Sequence 5' → 3'                     | 3' label | Oligo Type        |
|-----------------------|----------|--------------------------------------|----------|-------------------|
| CSF1R-I792T-UNI_F     | n/a      | <b><u>GCGCTTTCTTCAGTGCATCC</u></b>   | n/a      | Universal Forward |
| CSF1R-I792T-UNI_PROBE | FAM      | <b><u>CCGGGACGTAGCAGCTCGAAAC</u></b> | ZEN/IBFQ | Universal Probe   |
| CSF1R-I792T-WT_R      | n/a      | <b><u>CCAGTCCAAAGTCCCCAATC</u></b>   | n/a      | Wild type Reverse |

## CSF1R-I792T-MUT1 assay (FAM labelled)

TGGTCCTCAGGCCTCAGGGAAGGATAAACTGACTAATAATCTCTCT**GCGCTTTCTTCAGTGCATCCAC**  
**CGGGACGTAGCAGCTCGAAAC**GTGCTGTTGACCAGCGGACATGTGGCCAA**aAcTGGGGACTTTGG**  
**ACTGG**CTAGGGACATCATGAATGACTCCA**ACTATGTTGTCAAGGGCAATGTGAGTGCCGAGAGAGA**

Lower case letters denote bases changed in the mutant allele.  
 Probe sequence is in bold and shaded grey.  
 Primer sequences are in bold and underlined.

| Oligo CSF1R-I792T     | 5' label | Sequence 5' → 3'                     | 3' label | Oligo Type        |
|-----------------------|----------|--------------------------------------|----------|-------------------|
| CSF1R-I792T-UNI_F     | n/a      | <b><u>GCGCTTTCTTCAGTGCATCC</u></b>   | n/a      | Universal Forward |
| CSF1R-I792T-UNI_PROBE | FAM      | <b><u>CCGGGACGTAGCAGCTCGAAAC</u></b> | BHQ      | Universal Probe   |
| CSF1R-I792T-MUT_R     | n/a      | <b><u>CCAGTCCAAAGTCCCCAGTT</u></b>   | n/a      | Mutant Reverse    |



## Dot1l internal control (VIC labelled)

CTGATGGGTGTGGGCAGATCCTACAGAGTCCCATTGGCCACCATGTGTGCTACGCCTGAAATAAAGCCTT**GCC**  
**CCAGCACGACCATT**CAGGG**CCAGCTCTCAAGTCG**ACTGTAAGATGAAGCATAAGGATGCCAACTACTAACA  
GAAAACGACTAGAGGGGAAAAGAACAAGGAAACAGAAGACGCAGCACTCCGGCTTCCCTGGGTTGGCCAGT  
CACCTATGA

| Oligo CSF1R-I792T | 5' label | Sequence 5' → 3'                       | 3' label | Oligo Type |
|-------------------|----------|--|----------|------------|
| Dot1l_Forward     | n/a      | <b><u>GCCCCAGCACGACCATT</u></b>        | n/a      | WT Forward |
| Dot1l_Probe       | VIC      | <b>CCAGCTCTCAAGTCG</b>                 | BHQ      | WT Probe   |
| Dot1l_Reverse     | n/a      | <b><u>TAGTTGGCATCCTTATGCTTCATC</u></b> | n/a      | WT Reverse |

Probe sequence is in bold and shaded grey  
Primer sequences are in bold and underlined

## DNA extraction method

DNA is extracted from ear clips using Applied Biosystems Taqman Sample-to-SNP Kit and qPCR run using 1:10 dilution from the crude preparation.

## qPCR master mix

1X

|   |          |
|---|----------|
| Applied Biosystems GTX Taqman master mix      | 5 µl     |
| Dot1l_Forward (20 µM)                         | 0.225 µl |
| Dot1l_Reverse (20 µM)                         | 0.225 µl |
| Dot1l_Probe (5 µM)                            | 0.2 µl   |
| FAM Assay (probe 5 µM & primers 15 µM each)   | 0.3 µl   |
| ddH2O   | 1.55 µl  |
| DNA (1:10 dilution of ABI Sample-to-SNP prep) | 2.5 µl   |

Each sample is ran in technical duplicate. Seven WT and/or mutant controls are also included in duplicate along with non-template controls.

## qPCR cycling conditions

qPCR instrument: Applied Biosystems 7500/7900 or ThermoFisher QuantStudio 7

95°C for 20 sec  
Then 40 cycles of;  
95°C for 3 sec  
60°C for 30 sec

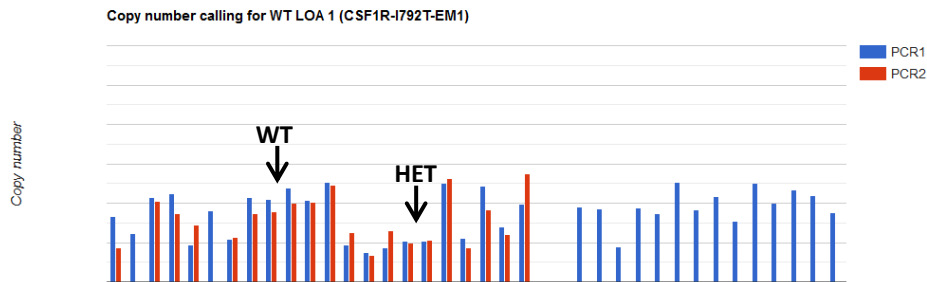
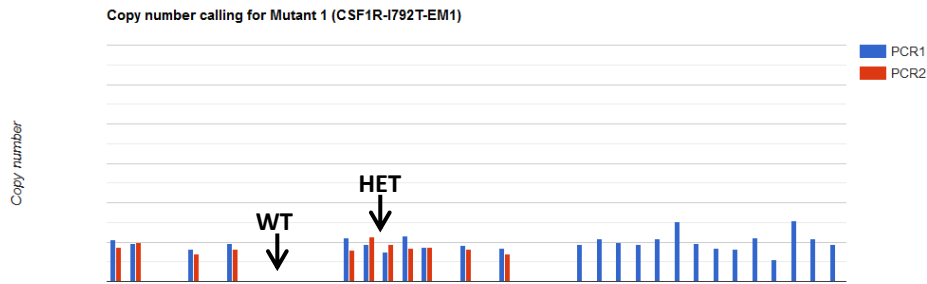




## Analysis

The results are analysed using CopyCaller software v2.0 from Applied Biosystems or in-house software that is based on CopyCaller v2.0.

CSF1R-I792T-WT1 and CSF1R-I792T -MUT1 assays copy called results, image showing copy number chart for WT and Mutant assays (Task 278187 results)



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Date: 11/06/2020

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