

**EMMA ID: 13580****Gene: *Bspf2*****Common name: *Bspf2-em1\_11*****Allele: *Bspf2<sup>em1(IMPC)Hmg</sup>***

## Allele Information

For more information on production,guides and mutation, search for gene/project, go to project summary, go to production plan, go to production outcome and "more details"

<https://www.gentar.org>

IMPC mouse phenotype data, search by the gene name

<http://www.mousephenotype.org/>

## Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice. For example: mutant positive, wild type positive = Heterozygous. In addition to the expected product, the mutant assay may also amplify the endogenous wild type sequence, which will appear as a larger band on an agarose gel. The presence of this extra band will depend on the size of the original deletion.

### PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Wild type	Bspf2_2G for	Bspf2_3rev 200	948/1558
Mutant	Bspf2_3for 100	Bspf2_3rev 200	429

### Primer sequences

Primer Name	Sequence 5' --> 3'
Bspf2_3for 100	caccatgatagactgtgtccatc
Bspf2_3rev 200	CTGGTTTCACTCCCTACTTGTCAT
Bspf2_2G for	tgaacactgttctcagattaatccct

### PCR setup (Qiagen, Hot Start Plus)

Component	Volume ( $\mu$ l) 1x	Final conc.
DNA (~ 50-100 ng)	2	
Q-Solution (5x)	2,5	0,5
PCR-Buffer (10x)	2,5	1
DNTP mix (10 mM)	0,5	0,2
MgCl <sub>2</sub> (25 mM)	1,5	1,5
Primer 1 (10 pmol/ $\mu$ l)	1	0,4
Primer 2 (10 pmol/ $\mu$ l)	1	0,4
Taq Polymerase (5 U/ $\mu$ l)	0,3	0,06
H <sub>2</sub> O*	13,7	
Final volume	25	

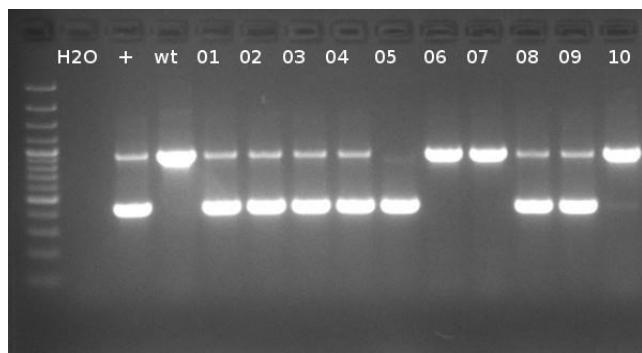
\* The amount of H<sub>2</sub>O is adjusted with the number of primer.

### Amplification conditions

PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C 68-58 ( $\downarrow 1^{\circ}\text{C/Cycle}$ ) 72°C	30 sec 45 sec 1.5 min	39
3 Polymerisation	72°C	15 min	1
4 Cooling	4°C	hold	1

**Touch-Down cycling protocol:** first 10 cycles anneal at 68°C, decreasing 1°C per cycle, next 30 cycles anneal at 58°C.  
 These PCR conditions have been optimized for our methods and preparation kits. Adoptions may be required.

### Gel Image



Separated by gel electrophoresis on a 2% agarose gel.