



Table of contents

Table of contents .....	1
1. Schematic representation of the locus.....	2
1.1. Overview .....	2
1.2. Strategy chosen: flox of exons 5 and 6.....	3
2. Construct used for homologous recombination in ES cells: Nr3c2 project.....	4
2.1. Legend .....	4
2.2. Map of targeting vector plasmid.....	4
2.3. 5' homology arm (4.1 kb).....	5
2.4. Floxed fragment (2.3 kb) .....	6
2.5. PGK-Neo region .....	6
2.6. 3' homology arm (3.2 kb).....	7
2.7. Vector backbone sequence .....	7
3. ES cell lines targeted and validation data .....	9
3.1. ES cell lines targeted .....	9
3.2. PCR data on positive clone.....	9
3.3. Southern data on positive clone.....	11
4. Data on conditional and knock-out animals.....	15
4.1. Genotyping protocol and data.....	15
4.2. Evaluation of lethality of homozygote KO (KO/KO) .....	18

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This protocol has been prepared by Claudia Caradec, Engineer

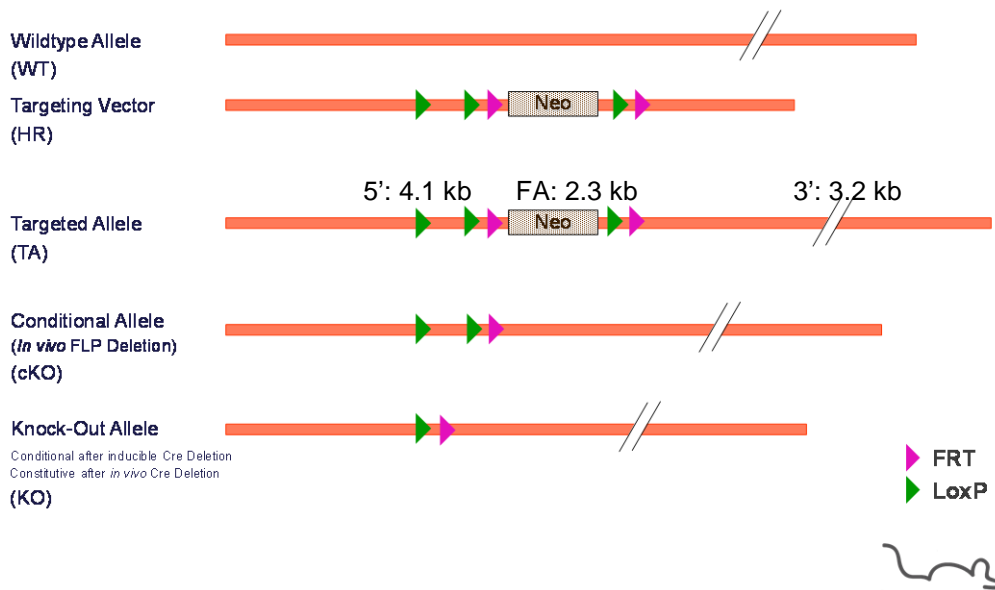
This protocol has been validated by Sylvie Jacquot, Ph.D., Project Manager

1. Schematic representation of the locus

1.1. Overview



## Overview Targeting Strategy



Legend:

5': 5' homology arm; FA: floxed fragment; 3': 3' homology arm  
 This schematic representation is not on scale

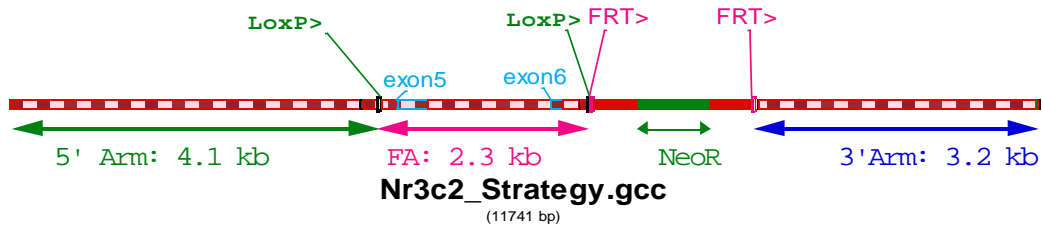


### 1.2. Strategy chosen: flox of exons 5 and 6

Nr3c2 gene (also named MR) is a member of the nuclear receptor family. Additional information on this gene can be accessed at

<http://www.informatics.jax.org/javawi2/servlet/WIFetch?page=markerDetail&key=15265>

#### Strategy used to generate the conditional knock out model

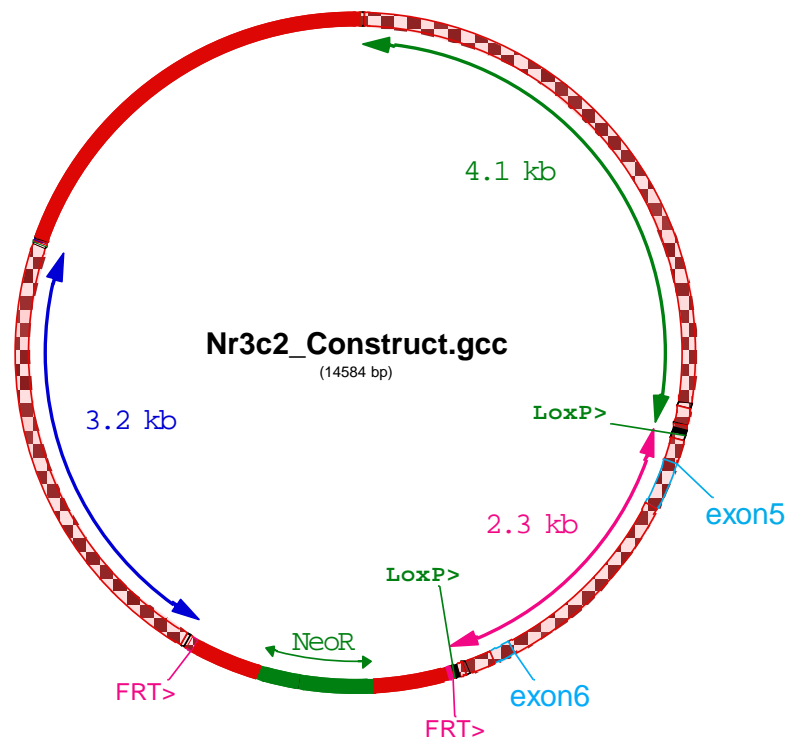


2. Construct used for homologous recombination in ES cells: Nr3c2 project

**2.1. Legend**

loxP sites are indicated in green ; FRT sites are indicated in purple; *Mus musculus* sequences are indicated in uppercase ; exogenous sequences are marked in lowercase.  
 The targeting vector was generated in 129Sv/Pas and was not fully sequenced

**2.2. Map of targeting vector plasmid**







**2.4. Floxed fragment (2.3 kb)**

ggccggccataaacttcgtataatgtatgctatacgaagttat ttaattaaATAGTACTGACCTGGGAGCTTGGCC  
 ATTTACCTTAAACTATTTTTGAAAGTAATCCTTTTTGTACAGGAGTTTTCTTAGGAGAGAGCCTTCTTGATTGCAAC  
 TCAATGGCTTGTGGAAGTCAGTTGTCCCCATGCAATGTGTTTCCGAAATGATATGTGCGTTTGCCTTATCCTTTCT  
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 AGCCCCACCGCCACCACCCAGAGCCCAGAAGAGGGGACCACATACATTGCTCCTACCAAGGAGCCATCAGTGA  
 ACTCTGCGCTGGTCCCGCAGCTCGCCTCGATCACGCGTGCCTCACGCCATCCCCGTCATGATCCTGGAGAACA  
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 CAGGCTCCGCCTGACAGCGGGCCCTGTTGTCCGCTTTGCATAGCATCCTGGCATGAGTACCGAAGACTCATGGCG  
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 CACTCTAATCAACTCTTCTGGTTAATAAATAAATAAATCCTGCTTGGGACTCAAGAGATGGTGGGGAAAGTACCT  
 TCATACAAACTGAGTTTGGATCCTCAGAACTGACAATTACATTACATATGGTCCCCATCTCAGAGAAGTGGCAGG  
 CCTGTAAACCCACCCTGGAGAGAGACAGAGACAGAGACAGAAGGATCCCTGGACTCAGGCCAGACAGCCTAGC  
 TCCACGTTCTGAGAAAGACCCGGCCTCAAAGATAAAGTAGACAATGATAGACAAGCCACGCCACATATGGCTT  
 CTCCATACTGCACCCACATGTCCGCACCTATACCTGTGCATGCACATGCATACACTTGCACATATGTCACAGGCA  
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 GTATCCCAGGCAACGGGGACCACAGAGTCAAAAAGACTAAAGTGGTCTCTGCTTTCAGACATGCAGATGGTGAC  
 AGTTAACCCCCCACCACCCATCCAAGTCAGTGTGTTGATTGTGTGGGATGAAGCAAGTGTACAGGTGACTG  
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 TAAGTGTCTTTCCGTGATTAATTGTTCTGTCTTAGGATTTAAAAACTTGCCTCTTGAGGACCAAATFACCCTC  
 ATCCAGTATTTGGATGTGTCTATCATCGTTTGCCTTGGATGAGATCGTACAAACATACGAACAGCCAATTT  
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 TATCAATAACAAATCAGTATATGTAACCTGTTACATCATGTATGTGGAACTGTCCCAAGAGAGAGAGcaccgggtg  
 ataacttcgtataatgtatgctatacgaagttat

**2.5. PGK-Neo region**

gcccggcgaagttcctattctctagaaagtataggaacttcgcccgaattctaccgggtaggggagggcgcttt  
 tcccaaggcagctcggagcatgcgcttagcagccccgctggcacttggcgctacacaagtgccctctggcctcg  
 cacacattccacatccaccggtagcgcgaaccggctccgctctttgggtggcccttcgcgccacctctactcct  
 cccctagtcaggaagttccccccgccccgcagctcgcgctcgtgcaggacgtgacaaatggaagtagcacgctctc  
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aagtataggaacttcccgcgatccatcgacccccctgcagg

### 2.6. 3' homology arm (3.2 kb)

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TTTGTAACTGTACACAGCTGCTCAATGTGTTTTCTGACAATCATAATTACCCTTCCTTTTCGATCATAAAAGAA  
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AGACAGACCACAGTTTATGCAAAGGATTGAGGACTTTCATGCACACTAGGCAAAAGCATCTAGAAGTGAACCACAC  
ACCCAGCTCTTAGTAGTATTTTTAT

### 2.7. Vector backbone sequence

ggccactgagggcgcgatcgcaagcttatcgataccgctcgacctcgagggggggcccggtacccaattcgcccta  
tagtgagtcgtattacgcgcgctcactggccgctggtttacaacgctcgactgggaaaaccctggcggtaccca  
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ccaacagttgcgcagcctgaatggcgaatgggagcgcgcccctgtagcggcgcatgaagcgcggcggtgtggtggt



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gcacccaggctttacactttatgcttccggctcgtatgttgggtggaattgtgagcggataacaatttcacaca  
ggaaacagctatgacctgatctacgccaagcgcgcaattaaccctcactaaaggggaacaaaagctggagctcgcg  
gccgcggcgcgc



### 3. ES cell lines targeted and validation data

#### 3.1. ES cell lines targeted

The targeting vector was electroporated in P1 ES cells [MCI-129Sv/Pas background]

Number of clones screened: 372

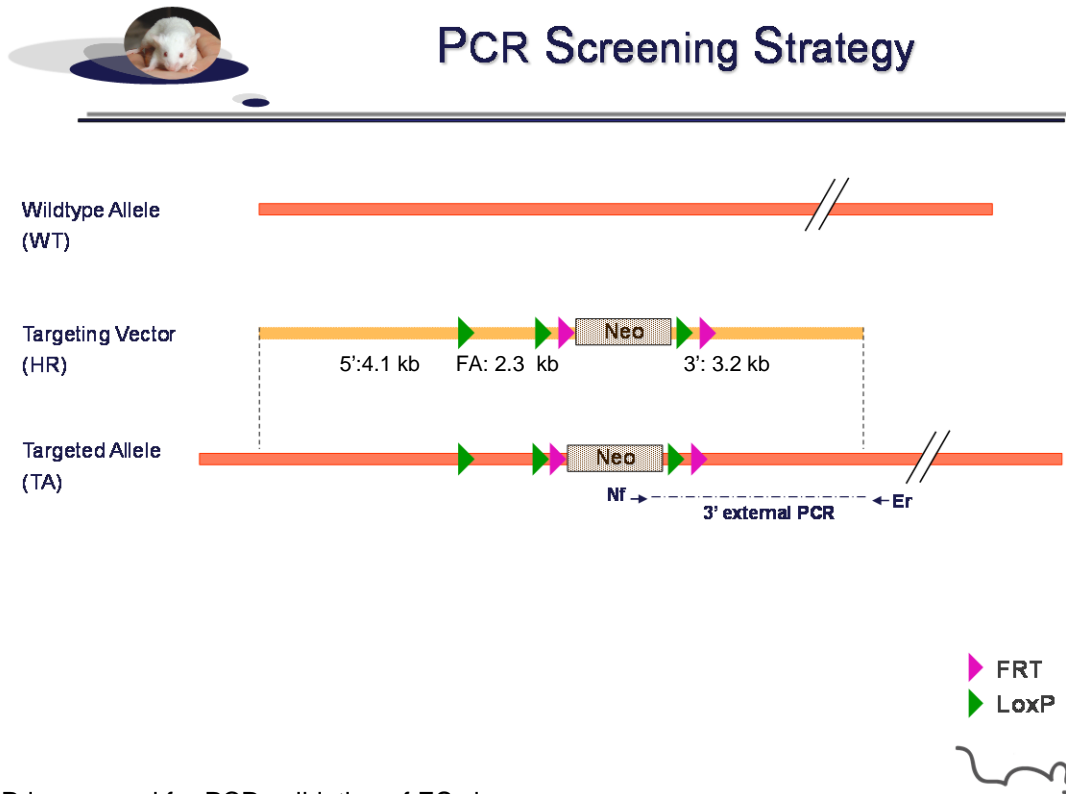
Number of positives: 3

Reference of clone used to generate the mouse line:

- clone **K179-30**

#### 3.2. PCR data on positive clone

##### 3.2.1. PCR screening strategy



Primers used for PCR validation of ES clone

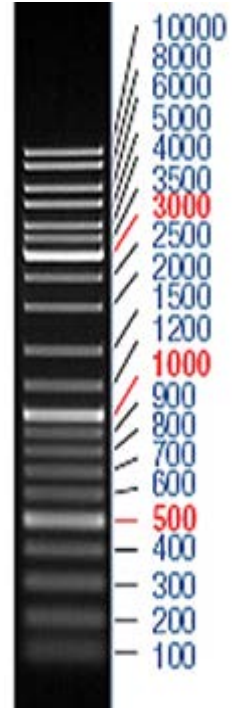
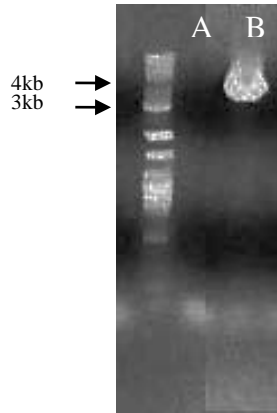
PCR	Primer Name	Primer sequences	PCR product size
3' external	Nf	CACGAGTAACTTCATGTGTGAACC	4.0Kb
	Er	AGGGGCTCGCGCCAGCCGAAGTGT	

### 3.2.1. Picture of PCR on positive clone

3' external PCR

ladder

**A:** WT clone  
**B:** positive clone

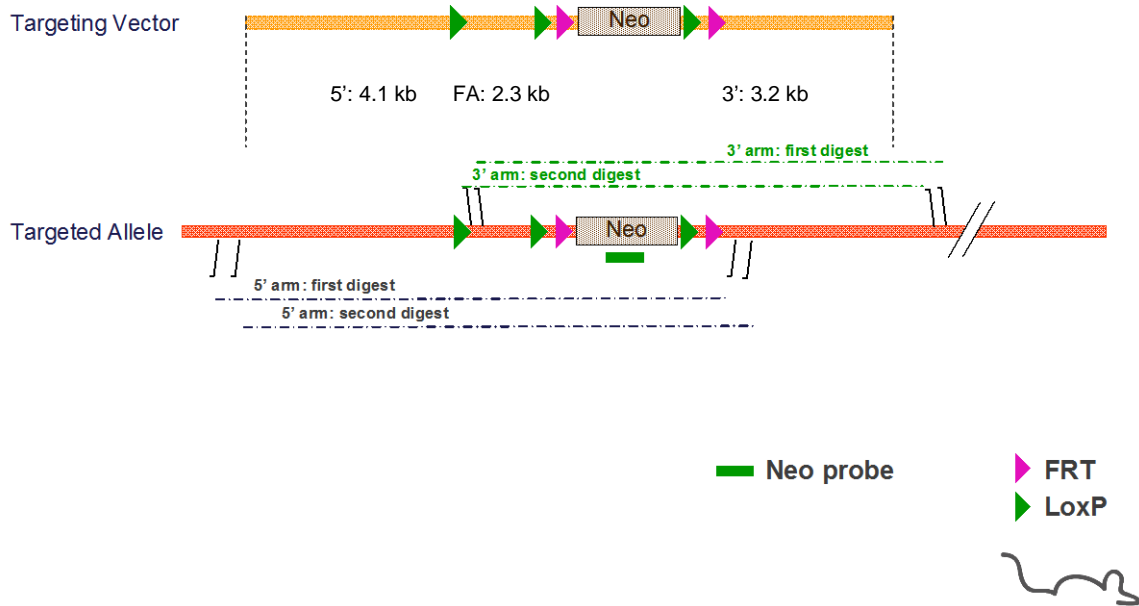


### 3.3. Southern data on positive clone

#### 3.3.1. Neo Southern strategy



## Southern Screening Strategy



Digestions used to validate the insertion

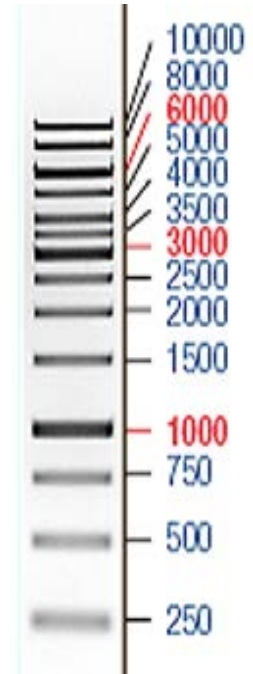
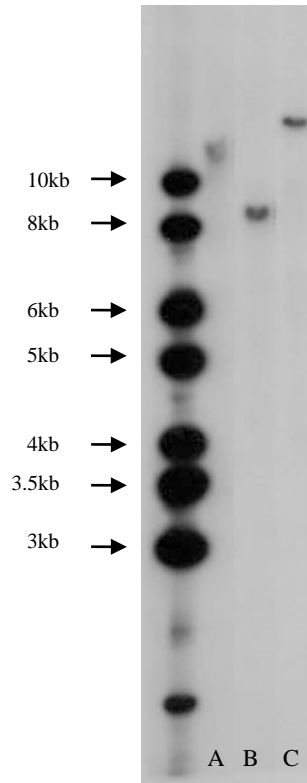
Probe	Name	Genomic DNA digest	WT allele (kb)	Targeted Allele (kb)
Neo	3' arm first digest	KpnI	/	12.4
	3' arm second digest	Afl II	/	8.7
	3' arm third digest	Hind III	/	14

### 3.3.2. Picture of Neo Southern

Neo southern blot: 3' arm validation

ladder

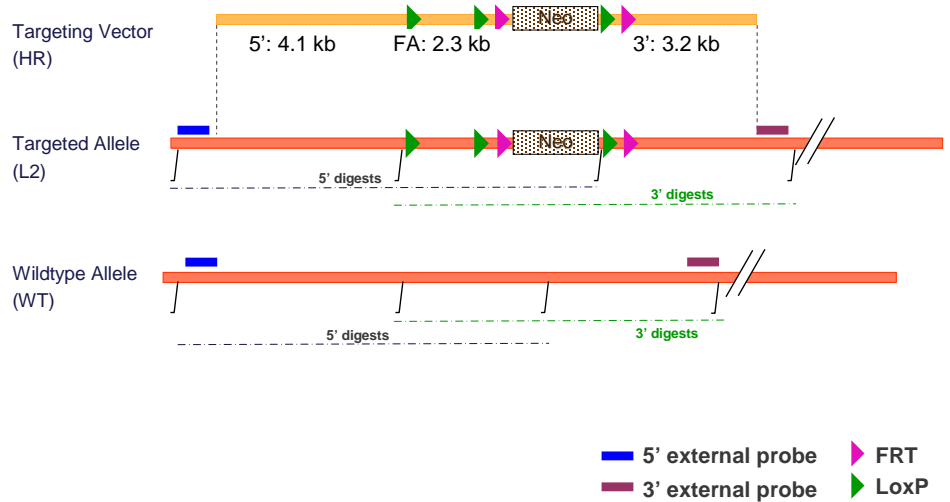
**A:** Kpn 12.4kb  
**B:** AflIII 8.7kb  
**C:** HindIII 14kb



### 3.3.3.External Southern strategy



## Southern Screening Strategy



Digestions used to validate with 5' probe

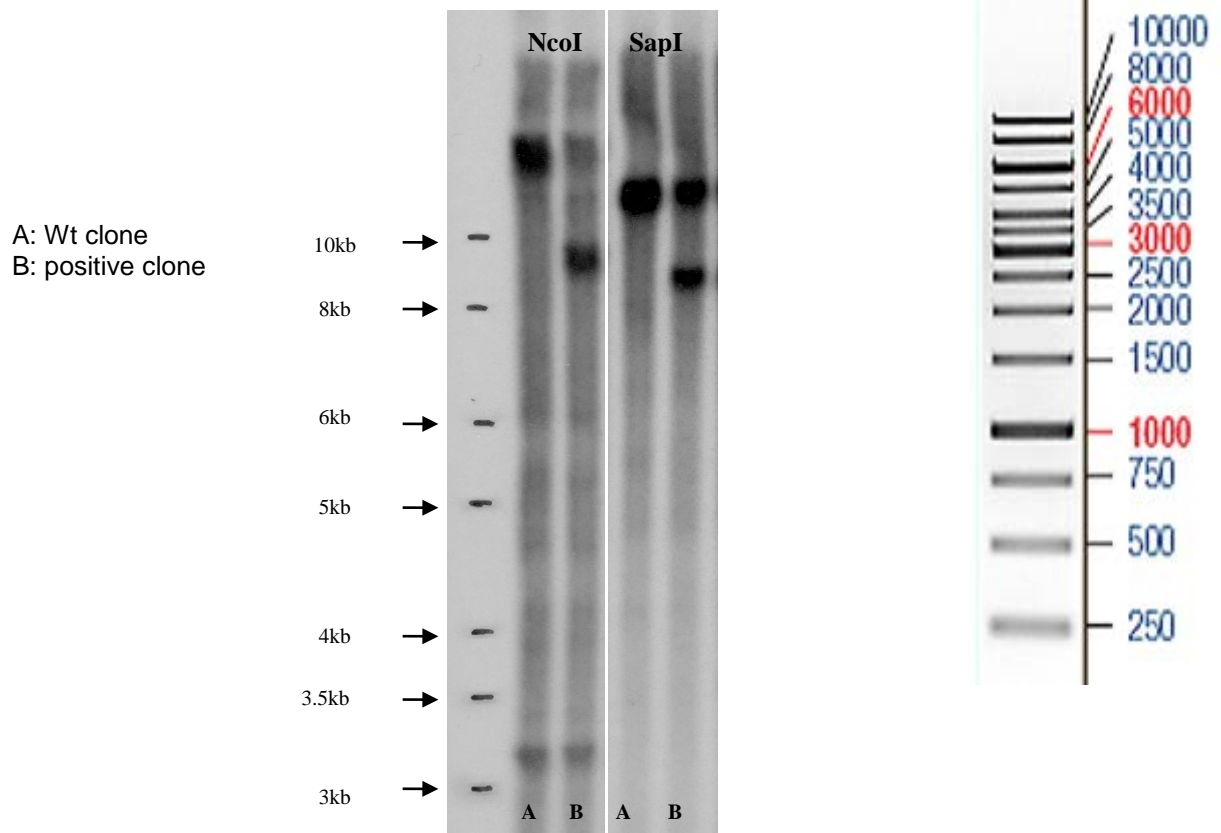
Probe	Name	Genomic DNA digest	WT allele (kb)	Targeted Allele (kb)
5' external	first digest	Nco I	15.6	9.6
	second digest	Sap I	12.7	8.9

Primers for the 5'probe synthesis:  
 CAGAACTGTGGCCTAGTCCACTTCCA  
 GCATCTCCCACTACAGTTTCAGGTGT

**3.3.4. Picture of Southern with external 5'**

5' external probe

ladder

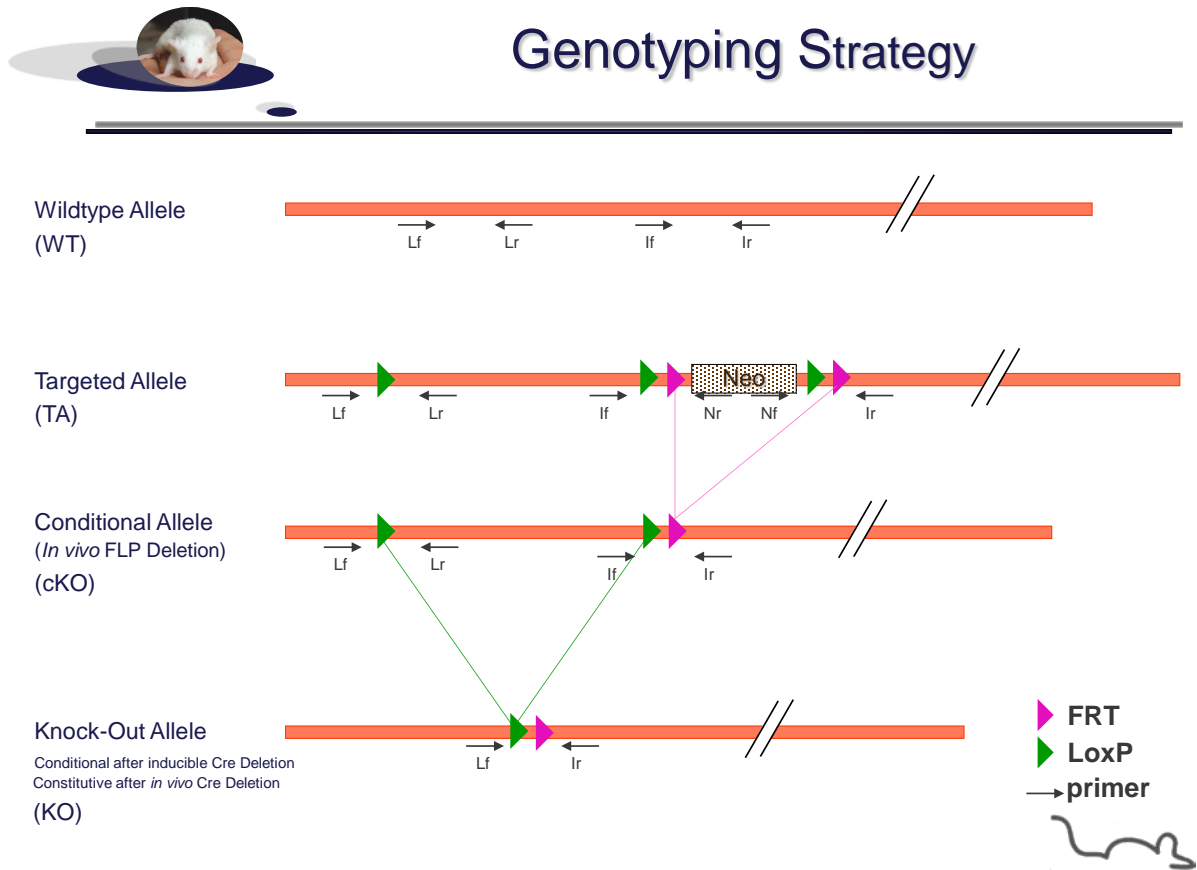


4. Data on conditional and knock-out animals

Both conditional and knock-out mouse models were backcrossed in C57BL/6J background.

4.1. Genotyping protocol and data

4.1.1. Genotyping strategy



Sequence of primers used for genotyping

Position	Primers	Sequence
Lf	783	CCACTTGTATCGGCAATACAGTTTGTGTC
Lr	785	CACATTGCATGGGGACAACACTGACTTC
Ef	786	GGAGATCGTACAAACATACGAACAGC
Er	788	CTGTGATGCGCTCGGAAACGG



PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer (see the map above)	Conditional allele (cKO)	Knock-Out allele (KO)	WT allele (WT)
Presence of the distal 5'loxP	783-785	Lf / Lr	358	---	308
Excision of the selection marker	786-788	Ef / Er	565	---	464
Total Excision (excision of the floxed exon(s), i.e. knock out)	783-788	Lf / Er	2812*	454	2652*

\* This PCR product will not be observed using our PCR genotyping conditions (see description below)  
 --- No Amplicon should be obtained

#### 4.1.2.PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:	Volume:
-10x Buffer (Roche)	2.5µl
-dNTPs 10mM (Amersham Biosciences)	0.5µl
-Taq DNA Polymerase (Roche)	0.2µl
-DNA (50ng/µl)	3µl
-5' primer (100 µM)	0.125µl
-3' primer (100 µM)	0.125µl
-Sterile H2O	up to 25 µl

Cycling conditions:

Temp	Time	#Cycles
94°C	3min	1
94°C	1min	2
62°C	1min	
72°C	1min	
94°C	30s	30
62°C	30s	
72°C	30s	
72°C	3min	1
4°C	∞	

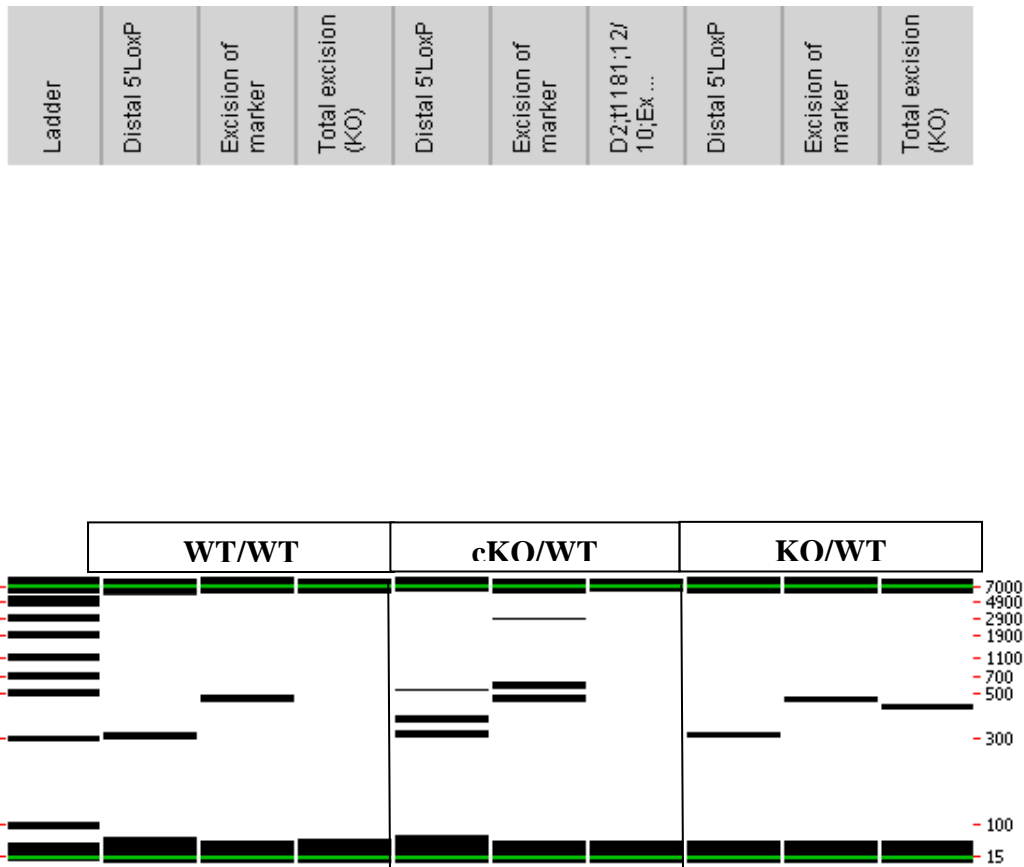
**NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.**



**4.1.3. Picture of genotyping with various alleles**

Analysis of PCR products pattern was not done by gel electrophoresis but using LabChip® 90 microfluidic apparatus. PCR products were run on the HT DNA 5K LabChip® 90 Assay Kit.

Representative genotyping picture



Note that as this technology is more sensitive than gel analysis, non specific signals and/or primer dimers may be visible on the picture.

#### 4.2. Evaluation of lethality of homozygote KO (KO/KO)

Males knock-out heterozygotes (KO/WT) were crossed with females knock-out heterozygotes (KO/WT). Offspring was genotyped to evaluate the ratio of the different genotypes. Results are provided in the table below.

Genotype	WT/WT	KO/Wt	KO/KO	Total
<b>Number of pups obtained</b>	24	33	1	<b>58</b>
<b>Experimental Ratio</b>	41,4%	56,9%	1,7%	<b>100%</b>
<b>Theoretical Ratio</b>	25%	50%	25%	<b>100%</b>
<b>Theoretical Ratio if KO/KO are not viable</b>	33%	66%	0%	<b>100%</b>

The Nr3c2a1 knock-out homozygotes are subviable.

#### Legend:

- >13% Homozygous = Viable
- >0% and ≤13% = Subviable
- 0% = Lethal