



Genotyping protocol

Dnm1

IR00004152 / P4152

(ICS internal reference)

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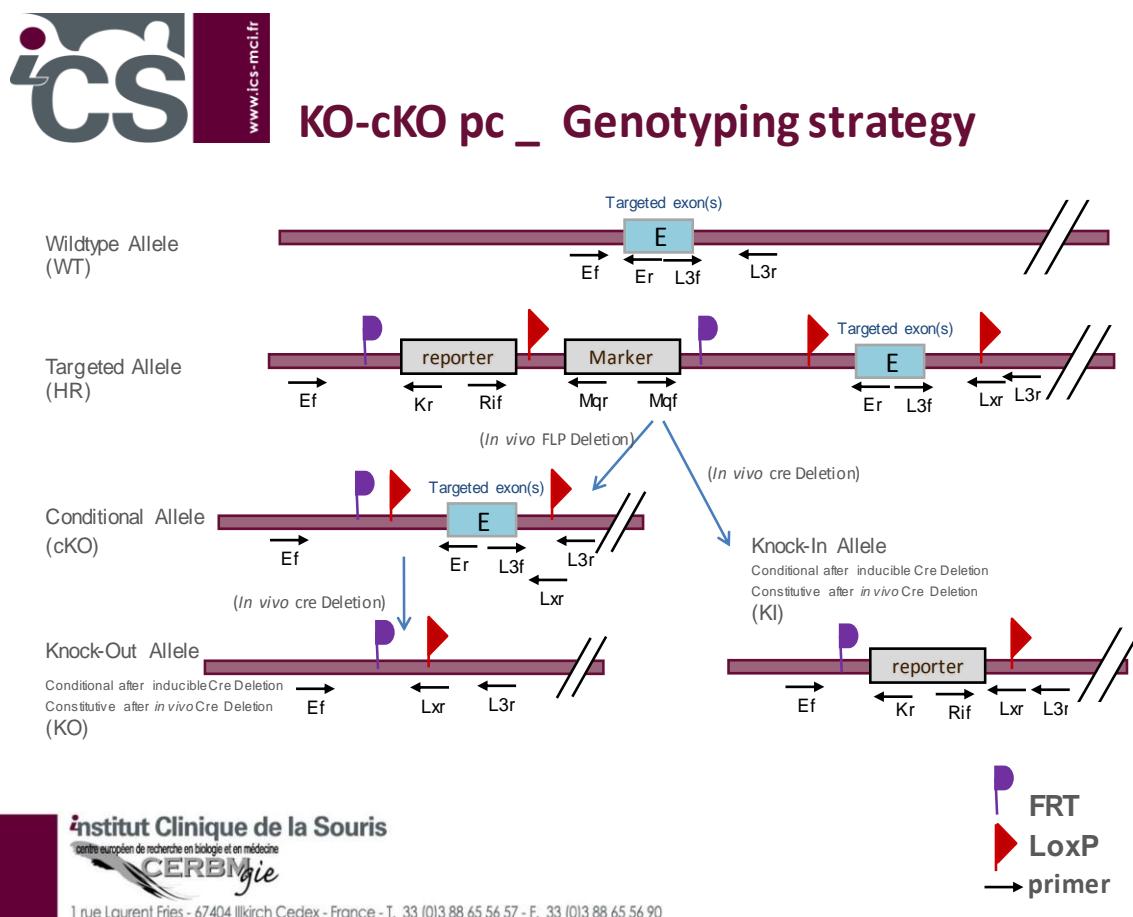
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1. Genotyping protocol and data

This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **Dnm1l** Constitutive Knockout / Conditional Knockout (KO-cKO x Cre) project.

1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	7544	GATTGGGCAGGAGCAAGATTCTC
Er	7548	GTGAGCTGACTGATGTAGGTGCTG
Kr	3278	GGGCAAGAACATAAAGTGACCCCTCC
L3f	7546	GCTCATATAAACATGCAGATGCAC
L3f ²	7545	CCTCTTGTCTCCAAGGATACCG
L3r	7547	GCAGCAATATAAGCATTAGAACAGG
Lxr	3255	ACTGATGGCGAGCTCAGACCATAAC
Rif	5966	GCACATGGCTGAATATCGACGGT

²: for a selected position, a second primer was designed

PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer <i>(see the map above)</i>	Targeted allele (HR)	conditional allele (cKO)	KI allele	WildType allele
5' part of the selection marker	7544-3278	Ef / Kr	312	---	312	---
Presence of the distal loxP	7546-7547	L3f / L3r	246	246	---	180
Distal loxP specific PCR	7545-3255	L3f ² / Lxr	216	216	---	---
Excision of the selection marker	7544-7548	Ef / Er	7329*	425	---	268
Cre total excision	5966-3255	Rif / Lxr	3382*	---	471**	---

*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

**: this PCR is only verified if mice are generated

---: no Amplicon should be obtained

1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:

- FastStart PCR Master (Roche)
- DNA (50ng/ μ l)
- 5' primer (100 μ M)
- 3' primer (100 μ M)
- Sterile H₂O

Volume:

- 7.5 μ l
- 1.5 μ l
- 0.06 μ l
- 0.06 μ l
- up to 15 μ l

Cycling conditions:

Temp	Time	#Cycles
95°C	4min	1
94°C	30s	
62°C	30s	34
72°C	1min	
72°C	7min	1
20°C	5min	1

NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.

2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Héault Y, Pavlovic G.
Genesis. 2012 Jun;50(6):482-9. doi: 10.1002/dvg.20826. Epub 2012 Mar 20.