

EMMA health monitoring procedures

Brief description of housing system, health monitoring programme and health status of the EMMA SPF live colonies

Type of facility

Animals distributed by EMMA are bred in SPF (Specific Pathogen Free) barriered facilities in which all materials are sterilized before entry. Staff entering the barriered areas must shower and change into clean unit clothing. Where appropriate, staff working within the units is also required to wear, gloves, face masks, mob caps and over shoes.

Housing system

Animals are maintained in either flexible film isolators or IVCs (Individually Ventilated Cages) or in conventional cages in barriered areas under positive pressure and are given autoclaved bedding, autoclaved or irradiated food and filtered or chlorinated water. Animals reared in IVCs are cage changed under laminar flow hoods.

Sentinel programme

The health status of each animal room is monitored on a regular basis e.g. 4 times per year when mice are reared in IVCs or monthly when mice are bred in conventional cages in barriered areas. These screening programmes involve exposing sentinel animals to dirty bedding collected from other IVCs within the mouse room. Some colonies e.g. those reared in isolators are sampled directly.

Health report

Before receiving any mice from EMMA you will be sent a recent (< 3 months old) health report prepared in accordance with the FELASA (Federation of European Laboratory Animal Science) recommendations. This health report will give details of the agents tested, the number of animals tested and the analytical methods used.

The following pages provide a sample health report from the EMMA node that distributes the strain you are interested in. Note that this is a **sample** health report and **not a current report**. Current reports will be provided upon request. Additional specific health checks (beyond tests recommended by FELASA) are possible if required by customers for importation but will be charged to the customer. If you require any further information please contact the archiving/distribution centre handling your request.

HEALTH MONITORING REPORT

Health monitoring is conducted according with FELASA recommendations for the health monitoring of mouse, rat, hamster, guinea pig and rabbit colonies in breeding and experimental units (Lab Anim, March 4, 2014)

Institute	Helmholtz Zentrum München German Research Center of Environmental Health							
Address	Ingolstädter Landstr. 1 85764 Neuherberg							
Area	ICE Unit	Q1 / 2019	Q2 / 2019	Q3 / 2019	Q4 / 2019			
Rooms:	203,205,219,220	Date: 11.03.2019	Date: 11.06.2019	Date: 13.09.2019	Date: 11.12.2019			
Housing:	IVC							
Species:	Mouse							
Viruses		EAD result	EAD result	EAD result	EAD result	Laboratory	Method	
Mouse hepatitis virus	(MHV)	-	-	-	-	Charles River	PCR	
Mouse rotavirus	(EDIM)	-	-	-	-	Charles River	PCR	
Murine norovirus	(MNV)	-	-	-	-	Charles River	PCR	
Minute virus of mice	(MVM)	-	-	-	-	Charles River	PCR	
Mouse parvovirus	(MPV)	-	-	-	-	Charles River	PCR	
Theiler's murine encephalomyelitis virus	(TMEV)	-	-	-	-	Charles River	PCR	
Lymphocytic choriomeningitis virus	(LCMV)	-	nt	-	nt	Charles River	PCR	
Mouse adenovirus type 1	(FL)	-	nt	-	nt	Charles River	PCR	
Mouse adenovirus type 2	(K87)	-	nt	-	nt	Charles River	PCR	
Mousepox (ectromelia) virus		-	nt	-	nt	Charles River	PCR	
Pneumonia virus of mice	(PVM)	-	nt	-	nt	Charles River	PCR	
Reovirus type 3	(REO3)	-	nt	-	nt	Charles River	PCR	
Sendai virus	(SV)	-	nt	-	nt	Charles River	PCR	
Bacteria								
<i>Helicobacter</i> spp.		-	-	-	-	Charles River	PCR	
<i>H.hepaticus</i>		-	-	-	-	Charles River	PCR	
<i>H.bilis</i>		-	-	-	-	Charles River	PCR	
<i>H.typhlonius</i>		-	-	-	-	Charles River	PCR	
<i>Rodentibacter pneumotropicus</i> , <i>R. heyltii</i>	(formally <i>Pasteurella pneumotropica</i>)	-	-	-	-	Charles River	PCR	
Streptococci β-haemolytic (not group D)		-	-	-	-	Charles River	PCR	
<i>Streptococcus pneumoniae</i>		-	-	-	-	Charles River	PCR	
<i>Citrobacter rodentium</i>		-	nt	-	nt	Charles River	PCR	
<i>Clostridium piliforme</i> (Tyzzer's disease)		-	nt	-	nt	Charles River	PCR	
<i>Corynebacterium kutscheri</i>		-	nt	-	nt	Charles River	PCR	
<i>Mycoplasma pulmonis</i>		-	nt	-	nt	Charles River	PCR	
<i>Salmonella</i> spp.		-	nt	-	nt	Charles River	PCR	
<i>Streptobacillus moniliformis</i>		-	nt	-	nt	Charles River	PCR	
Parasitological infections								
<i>Aspicularis</i> sp.		-	-	-	-	Charles River	PCR	
<i>Syphacia</i> sp.		-	-	-	-	Charles River	PCR	
Coccidia		-	-	-	-	Charles River	PCR	
<i>Giardia</i> sp.		-	-	-	-	Charles River	PCR	
<i>Spiroplasma muris</i>		-	-	-	-	Charles River	PCR	
<i>Trichomonas</i> sp.		-	-	-	-	Charles River	PCR	
Others		-	-	-	-	Charles River	PCR	
Ectoparasites		-	-	-	-	Charles River	PCR	
Pathological lesions:								
none observed		"- " = negative result; "+" = positive test result; "nt" = not tested						

Comment: Health monitoring based on exhaust air dust (EAD) PCR analysis; additional tests of embryo transfer recipients

Embryo transfer recipients were tested negative.

Presence of opportunistic organisms will be communicated upon request.

Neuherberg,
19.12.2019

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